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Suboptimal *Trichomonas vaginalis* Antigen Test Performance in a Low-Prevalence Sexually Transmitted Infection Community

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**Trichomonas vaginalis** is the most common nonviral etiology of sexually transmitted infection (STI) worldwide (1). The OSOM Trichomonas rapid test (OSOM; Sekisui Diagnostics, San Diego, CA) is a rapid surrogate to microscopic analysis in symptomatic patients (2), but its performance in low-prevalence STI populations has been assessed on a limited basis in the literature (3). OSOM has widespread usage, as accreditation data from the College of American Pathologists report that over 300 participant laboratories utilize this assay on an annual basis (4). We sought to characterize the analytical and clinical performance of OSOM in a low-prevalence STI population on the basis of a commercial transcription-mediated amplification (TMA) reference.

Results from the performance of OSOM with 1,421 consecutive vaginal saline suspensions were audited from a 4-month interval in a low-prevalence southeastern Wisconsin STI population (5). The concomitant Aptima specimen transport tube from the health care encounter (98.7% endocervical, 1.3% vaginal), previously subjected to *Chlamydia trachomatis* and *Neisseria gonorrhoeae* TMA (Aptima Combo 2 assay; Hologic, San Diego, CA), was forwarded for retrospective *T. vaginalis* TMA (Aptima Trichomonas vaginalis assay; Hologic) evaluation on a TIGRIS direct tube sampling (DTS) system per the manufacturer’s specifications (6). A previous assessment of *T. vaginalis* TMA showed 100% molecular concordance between the data from vaginal saline suspension aliquots and endocervical specimens maintained in Aptima specimen transport tubes (7). In addition, Napierala et al. (8) have shown equivalent *T. vaginalis* TMA detection rates from endocervical swabs and vaginal swabs in our population. This study was governed by the Wheaton Franciscan Healthcare Institutional Review Board.

The low-prevalence STI population exhibited TMA detection rates of 6.4% and 0.6% for *C. trachomatis* and *N. gonorrhoeae*, respectively, while the *T. vaginalis* TMA detection rate was 4.0%. On the basis of a *T. vaginalis* TMA reference standard, the sensitivity and specificity of OSOM were 35.1% and 99.9%, respectively (Table 1). The kappa value for this data set was 0.502 (95% confidence interval, 0.346 to 0.658), and agreement was 0.973 (95% confidence interval, 0.963 to 0.981).

In an analogous fashion, 98 consecutive tandem vaginal saline suspensions and endocervical swabs from an increased-prevalence STI setting (*C. trachomatis* TMA detection rate, 11.2%; *N. gonorrhoeae* TMA detection rate, 6.1%; setting characterized previously [7, 9]) were retrospectively analyzed via OSOM and *T. vaginalis* TMA, respectively, per the manufacturer’s guidelines (2, 6). Improved concordance between OSOM and *T. vaginalis* TMA was observed within this study set, with a 21.4% *T. vaginalis* TMA detection rate (Table 2). An increased kappa value of 0.904 (95% confidence interval, 0.797 to 1.000) was calculated, with agreement of 0.969 (95% confidence interval, 0.907 to 0.992).

Campbell et al. (3) reported 94.7% sensitivity for OSOM compared to microscopy in a population with a 1.9% *T. vaginalis* detection rate. Molecular diagnostics did not factor into the determination of the overall *T. vaginalis* incidence. For a high-prevalence population, Huppert and colleagues (10, 11) reported 83 to 90% sensitivity for OSOM. One can hypothesize that the significant *T. vaginalis* TMA/*T. vaginalis* antigen discordance frequency in our low-prevalence population is related to organism burden (12) or to *T. vaginalis* TMA detection in asymptomatic patients. However, 56.8% of OSOM-negative/*T. vaginalis* TMA-positive data in the low-prevalence population were derived from symptomatic patients (determined by chart review to have pelvic pain, abdominal pain, vaginal discharge, irritation, or itching). This rate of symptom incidence did not differ from that for patients yield-

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**TABLE 1** Performance of OSOM Trichomonas rapid test and Aptima Trichomonas vaginalis assay in a low-prevalence STI community

<table>
<thead>
<tr>
<th>OSOM Trichomonas rapid test result</th>
<th>No. of specimens with indicated result by Aptima Trichomonas vaginalis assay</th>
<th>Total no. of specimens</th>
</tr>
</thead>
<tbody>
<tr>
<td>Positive</td>
<td>20 Positive</td>
<td>21</td>
</tr>
<tr>
<td></td>
<td>1 Negative</td>
<td></td>
</tr>
<tr>
<td>Negative</td>
<td>37 Positive</td>
<td>1,400</td>
</tr>
<tr>
<td></td>
<td>1,363 Negative</td>
<td></td>
</tr>
<tr>
<td>Total</td>
<td>57 Positive</td>
<td>1,421</td>
</tr>
</tbody>
</table>

* Repeat Aptima Trichomonas vaginalis assay testing yielded a negative result.

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**TABLE 2** Performance of OSOM Trichomonas rapid test and Aptima Trichomonas vaginalis assay in an increased-prevalence STI community

<table>
<thead>
<tr>
<th>OSOM Trichomonas rapid test result</th>
<th>No. of specimens with indicated result by Aptima Trichomonas vaginalis assay</th>
<th>Total no. of specimens</th>
</tr>
</thead>
<tbody>
<tr>
<td>Positive</td>
<td>18 Positive</td>
<td>18</td>
</tr>
<tr>
<td></td>
<td>0 Negative</td>
<td></td>
</tr>
<tr>
<td>Negative</td>
<td>3 Positive</td>
<td>80</td>
</tr>
<tr>
<td></td>
<td>77 Negative</td>
<td></td>
</tr>
<tr>
<td>Total</td>
<td>21 Positive</td>
<td>98</td>
</tr>
</tbody>
</table>

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**References**


TABLE 3 Symptomatic status of Aptima Trichomonas vaginalis assay-positive patients from low- and increased-prevalence STI communities stratified by OSOM Trichomonas rapid test result

<table>
<thead>
<tr>
<th>OSOM Trichomonas rapid test result</th>
<th>Low-prevalence community</th>
<th>Increased-prevalence community</th>
</tr>
</thead>
<tbody>
<tr>
<td>Positive</td>
<td>15 (75.0)</td>
<td>16 (88.9)</td>
</tr>
<tr>
<td>Negative</td>
<td>21 (56.8)</td>
<td>3 (100.0)</td>
</tr>
</tbody>
</table>

\[a P = 0.17, \text{versus those with positive OSOM result; significance test of proportions.}\]
\[b P = 0.54, \text{versus those with positive OSOM result; significance test of proportions.}\]


