

11-1-1986

Charge Transfer-oxy Radical Mechanism for Anti-cancer Agents

Peter Kovacic

University of Wisconsin - Milwaukee

James R. Ames

University of Wisconsin - Milwaukee

Paavo Lumme

University of Helsinki

Hannu Elo

University of Helsinki

O. Cox

University of Puerto Rico, Rio Piedras

See next page for additional authors

Authors

Peter Kovacic, James R. Ames, Paavo Lumme, Hannu Elo, O. Cox, and Michael D. Ryan

Charge transfer-oxy radical mechanism for anti-cancer agents¹

P. Kovacic¹, J.R. Ames¹, P. Lumme², H. Elo², O. Cox³, H. Jackson³, L.A. Rivera³, L. Ramirez³ & M.D. Ryan⁴

¹Department of Chemistry, University of Wisconsin-Milwaukee, Milwaukee, WI 53201 USA,

²Department of Inorganic Chemistry, University of Helsinki, Vuorik 20, SF-00100, Helsinki,

Finland, ³Department of Chemistry, University of Puerto Rico, Rio Piedras, Puerto Rico 00931,

and ⁴Department of Chemistry, Marquette University, Milwaukee, WI 53233, USA

Summary: The proposal is advanced that anti-cancer drugs generally function by charge transfer resulting in formation of toxic oxy radicals which destroy the neoplasm. Electrochemical studies were performed with some of the main types of agents: iminium ions (adenine iminium from alkylating species, iminium metabolite of 6-mercaptopurine, nitidine, other polynuclear iminiums) and metal complexes (Pt(II)diaquodiammine-guanosine, copper salicylaloximes). Reduction potentials ranged from -0.4 to -1.2 V. Literature data for quinones are presented and radiation is discussed. Based on the theoretical framework, a rationale is offered for the carcinogen-anti-cancer paradox and the role of antioxidants.

More than two decades ago the oxy radical hypothesis for carcinogenesis was advanced (Brues & Guzman Barron, 1951; Holman, 1956; Harman, 1956). Shortly thereafter, the proposal was placed on a broader, more systematic foundation (Harman, 1962; Kovacic, 1959 and 1960). This approach received scant attention until fairly recent times which have witnessed ever increasing support from a variety of disciplines (Ames, 1983; Mason, 1982; Demopoulos *et al.*, 1980). In general terms, the comprehensive theory states that oxy radicals are implicated in the action of most carcinogens, arising as the end product of metabolic processes, usually via charge transfer (CT). Apparently, the highly reactive radicals subsequently attack cellular DNA, as well as other crucial constituents, resulting in transformation to the oncogenic state. Specific application has been made to alkylating agents, quinones, metal complexes, iminium ions, radiation, carbon tetrachloride, 4-nitroquinoline 1-

oxide, and inert bodies (Kovacic *et al.*, in press).

The initial inklings (Holman, 1956; Warburg *et al.*, 1957) that reactive oxy species may play a role in anti-cancer action was shortly followed by a better developed, more comprehensive approach (Kovacic, 1959). A baffling paradox of oncology is the well-known phenomenon that generally the substances which induce cancer are also antineoplastic. If the premise is valid that these agents cause cancer by producing excessive amounts of oxy radicals, it may well be that their ability to combat the condition is intimately related to the same chemical property. An essential component of the overall picture is the corollary that many tumor cells are more susceptible than normal ones to elevated concentrations of oxy radicals, thus providing the requisite specificity. Supporting evidence may be found from the early days of oncology (Kovacic, 1959), as well as newer data which will be presented in the discussion section.

Recently the suggestion was made that iminium species (1), usually in conjugated form, play important roles biologically in a

¹Presented in part at the 189th national meeting, American Chemical Society, Miami, FL, MEDI Abstracts, 81 (1985).

Correspondence: P. Kovacic.

variety of redox transformations (Kovacic, 1984). These entities might then function catalytically at the active site as electron conduits for the formation of superoxide, a precursor of other oxy radicals (Fridovich, 1983). This concept is now applied to the anti-cancer domain.

The principal objective of the present work was to determine the electrochemical characteristics of several main categories of antineoplastic agents: iminium ions (adenine iminium from alkylating species, iminium metabolite of 6-mercaptopurine, nitidine, other polynuclear iminiums) and metal complexes (Pt(II) diaquodiammine-guanosine, copper(II) salicylaloximes). Literature data for quinones and other CT agents are presented, and radiation is discussed. The results are treated within the context of the unifying theory for anti-cancer action involving CT with production of toxic oxy radicals. The carcinogen—anti-cancer paradox is addressed, as well as the role of antioxidants.

Materials and methods

Isoquinolinium salts **7** and **8** were obtained from Prof Mark Cushman (Cushman *et al.*, 1984). Literature methods were used for synthesis of purine-6-sulfinate **6** (mp 178°C (dec.), lit. (Doerr *et al.*, 1961); mp 175°C (dec.), 3-benzyladenine chloride **3** (mp 254–260°C with prior darkening, lit. (Abshire and Berlinquet, 1964; mp 261–267°C), 3-benzyladenine (3-HCl) (mp 268–270°C, with prior darkening, lit. (Abshire & Berlinquet, 1964; mp 284–287°C) and 1-methyladenosine iodide **4** (mp 190–195°C (dec.) (Jones & Robins, 1963). Elemental analyses were satisfactory for the compounds whose melting points differed appreciably from literature values. Benzo-thiazoloquinolinium salts **9** (Cox *et al.*, 1982 and unpublished results), and copper(II) salicylaloximates **11** (Lumme & Korvola, 1975; Lumme *et al.*, 1984) were prepared as described. *cis*- and *trans*-Diaquodiammine platinum(II) nitrates were obtained from the corresponding DDPs by stirring with two

equivalents of AgNO₃ in H₂O for 3 and 1 h, respectively, and filtering the AgCl precipitate (Marcelis *et al.*, 1980). The solution was evaporated to dryness in a vacuum over H₂SO₄ to yield the product. Complex formation was attempted (Dehand & Jordanov, 1976) with guanosine and the *cis*-diaquo reagent for 30 min, since this Pt compound is reported to be quite reactive (Marcelis *et al.*, 1980). However, no precipitate formed; the solution was evaporated under vacuum to furnish a solid material.

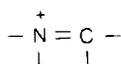
Cyclic voltammetry and polarography were performed on an ECO model 550 potentiostat with a PARC model 175 waveform generator. All solutions were degassed for 15 min with pre-purified dinitrogen that was passed through an oxygen scrubbing system. The working electrodes were a platinum flag or a hanging mercury drop (HMDE). Reference electrodes were an IBM aqueous Ag/AgCl or a Corning SCE both in saturated KCl. The counter electrode in all cases was a platinum wire. The supporting electrolyte was tetraethylammonium perchlorate (G.F. Smith Chemical Co.). The solvents, *N,N*-dimethylformamide and dimethyl sulfoxide were obtained from Aldrich Chemical Co. in the highest possible purity, in addition to *cis*-DDP, *trans*-DDP and guanosine hydrate. Buffer solutions of pH 3.3, 3.9 and 4.8 (HOAc/OAc⁻) (compound **6**) and pH 6 (50% ETOH/buffer, KHP) (compounds **7** and **8**) were used for cyclic voltammetry.

Results and discussion

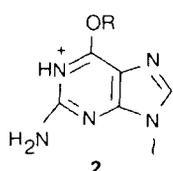
Iminium ions

1. Purines

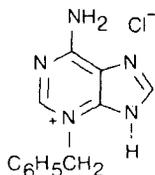
(a) *Alkylated DNA models*: The alkylating agent class contains a large group of antineoplastic agents, including nitrogen mustards, epoxides, aziridines, triazenes, *N*-nitroso compounds, and alkylalkanesulfonates (Reich, 1981). Some have progressed to the stage of practical use in chemotherapy. As is well established, the diverse types also



1



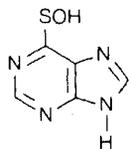
2



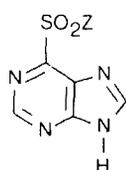
3



4

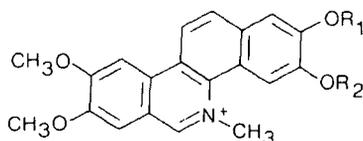


5

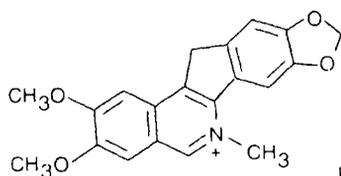


a) Z=H b) Z=Na

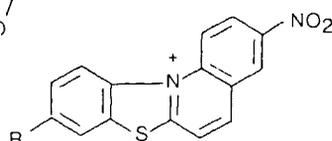
6

a) R₁+R₂=CH₂ b) R₁=H R₂=CH₃

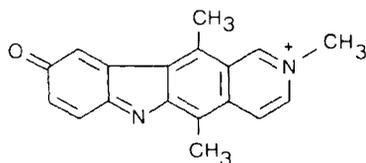
7



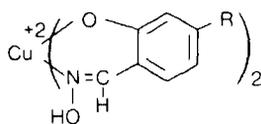
8

a) R=H b) R=OCH₃

9

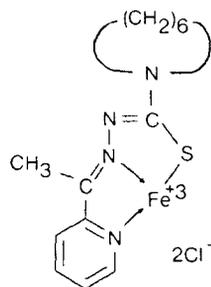


10



a) R=H b) R=OH

11



12

generally function as carcinogens (Miller & Miller, 1983). Concomitant production of oxy radicals has been observed with various members (Ames, 1983; Floyd, 1982). Although the precise role of these reactive intermediates has not been ascertained, it appears that DNA strand cleavage may be a crucial event (Floyd, 1982).

In a recent investigation of the mechanism of carcinogenesis, a novel proposal was advanced in which the salt form (iminium) of alkylated nucleic acid was assigned a key function as a CT agent (Kovacic *et al.*, in press). The purines (guanine and adenine) of DNA are the principal targets of attack (Miller and Miller, 1983). For example, the ionic

structure 2, a conjugated form of iminium (1) is generated from O-6 alkylation of guanine and could conceivably undergo one-electron reduction. Electrochemical data from the literature (Dryhurst, 1977) and our own studies (Kovacic *et al.*, in press) are in reasonable accord with the current picture relating site of alkylation and defect persistence to oncogenic response. Thus, it appears quite plausible that the salt form is functioning in a catalytic manner as a generator of toxic oxy radicals.

In order to test this concept as applied to anti-cancer alkylating agents, salts derived from alkylation of adenine and adenosine were investigated electrochemically as models. 3-Benzyladenine chloride 3 gives irreversible reduction values of about -1.0 V (Table I). Upon addition of strong base the potentials become more negative and the current drops until, with excess base, there is no reduction before the background current, due to generation of the nonreducible, nonionic base via loss of HCl. Occurrence of this transformation was confirmed by electrochemical studies on the free base which gave no reduction before background. A second model consisted of the nucleoside with the base alkylated at a different site, namely, N-1. For 1-methyladenosine iodide 4, the most

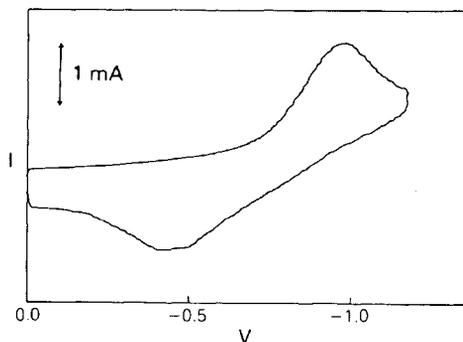


Figure 1 Cyclic voltammogram of 4 in DMF, Pt electrode, scan rate 100 mV/s.

positive figure obtained for the reduction potential was -0.96 V (DMF) (Figure 1, Table I). The results were less favorable in DMSO. The product of the reduction is probably a dimer, since coupling has been observed from electrolysis of purine bases in non-aqueous solvent (Yao *et al.*, 1976).

Our data are in agreement with previous investigations with adenine in aqueous acid. $E_{1/2}$ values, -1.05 to -1.07 V, were reported, which varied linearly with pH (Dryhurst, 1977). Adenosine and adenylic acid behaved similarly. Evidently salt formation occurs by preferential protonation at N-1 (Saenger, 1984).

There are several possible sites for adenine alkylation. The preferred one in vivo is generally the N-1 position. Reaction at N-7 is also commonly observed, whereas N-3 attack varies in degree (Shooter, 1972; Rajalakshmi *et al.*, 1982). Alkylation at any of these positions would produce a potential CT agent capable of catalytic operation. The N-3 position has been suggested as an important locale in the carcinogenic process (Lijinsky, 1976). It is significant that N-7 adenine salts possess physiological activity (Iio *et al.*, 1985).

(b) *6-Mercaptopurine*: The properties of this drug are summarized in Table VII. It, as well as related materials, is evidently converted to the corresponding nucleotide (Ishiguro *et al.*, 1984) via

Table I Cyclic voltammetry of N-alkylated adeninium and adenosinium halides.^a

Compound	[OH ⁻] mM	-E _p	
		DMF	DMSO
3-Benzyladenine chloride	—	0.96	1.00
	0.49	1.03	1.19
	0.99	— ^b	— ^b
3-Benzyladenine	—	— ^b	— ^b
1-Methyladenosine iodide	—	0.96	1.23
	0.49	0.96	1.23
	0.99	— ^b	— ^b

^a 100 mV/s, tetraethylammonium perchlorate (0.1 M), substrate (0.5 mM), Pt electrode, irreversible, vs. SCE.

^b No reduction of substrate before background reduction.

the nucleoside (Chabner, 1981; Christie *et al.*, 1984) followed by insertion into the DNA chain. The thiol appears to be oxidized to the unstable sulfenic acid 5 which undergoes further conversion to the isolable sulfinic acid 6a (Hyslop & Jardine, 1981; Nelson, 1982). The acids could exist in an ionic (iminium) form (cf. 2) either from intra- or intermolecular nuclear protonation.

Since an oxidative metabolite is thought to be the active agent, we obtained data on the reduction potential for 6. In DMF the E_p varies from -1.0 V for the iminium from nuclear protonation to > -2.0 V for the sodium salt (6b) (Table II). In aqueous buffer 6 exhibits potentials (V) that vary linearly with pH ($E_p = -0.44 - 0.100$ pH); the most positive value was -0.77 V (pH 3.3). Hence reduction is facilitated by increasing acidity. The results in aqueous media are in agreement with data (V) from an earlier study (Dryhurst, 1969) in which 6 exhibited $E_{1/2} = -0.37 - 0.094$ pH (pH 1–9.1). Reduction involved the N-1=C-6 bond giving the dihydro product (Dryhurst, 1977). Also included were the parent thiol, $E_{1/2} = -0.79 - 0.116$ pH (pH 0–5), purine 6-sulfonic acid, $E_{1/2} = -0.45 - 0.078$ pH (pH 1–7), and 6-purinyldisulfide, $E_{1/2} = -0.0$ V. From these findings Dryhurst concluded that

Table II Cyclic voltammetry of purine-6-sulfonic acid.^a

[Acid] mM	$-E_p$	
	DMF	H ₂ O ^b
—	>2.0	— ^c
HClO ₄	0.46	1.24
HClO ₄	0.91	1.03, 1.23
HClO ₄	1.3	1.00, 1.22
HOAc	pH 3.3	— ^c
HOAc	pH 3.9	— ^c
HOAc	pH 4.8	— ^c

^a 100 mV/s, tetraethylammonium perchlorate (0.1 M, DMF), 6b (0.5 mM), HMDE vs. SCE, irreversible.

^b HOAc/OAc[−] buffer.

^c Not examined electrochemically.

Table III Electrochemistry of fused derivatives of quinolinium and isoquinolinium salts.^a

Compound	$-E_p$			Electrode technique ^c
	DMF	DMSO	H ₂ O ^b	
7a	0.90	— ^d	— ^d	Hg, P
	1.07	1.09	— ^d	Pt, CV
7b	— ^d	— ^d	1.15	Hg, CV
	0.99	— ^d	— ^d	Hg, P
8	1.01	— ^d	0.89	Hg, CV
	1.11	1.14	— ^d	Pt, CV
	1.24	— ^d	— ^d	Hg, P
9a	1.25	— ^d	1.35	Hg, CV
	1.34	1.35	— ^d	Pt, CV
	0.39, 1.18	0.42, 1.12	— ^d	Hg, P
9b	0.42, 1.19	0.45, 1.17	— ^d	Hg, CV
	0.65, 1.21	0.64, 1.16	— ^d	Pt, CV
	0.42, 1.18	0.45, 1.12	— ^d	Hg, P
9b	0.45, 1.20	0.47, 1.16	— ^d	Hg, CV
	0.66, 1.17	0.67, 1.17	— ^d	Pt, CV

^a 100 mV/s, tetraethylammonium perchlorate (TEAP, 0.1 M), substrate (0.5 mM), vs. SCE.

^b Buffer (KHP) pH 6, no TEAP.

^c P—polarography, CV—cyclic voltammetry.

^d Not examined electrochemically.

5 should reduce at a value between those for 6b and the disulfide.

2. Fused derivatives of quinolinium and isoquinolinium salts. This class is represented by the alkaloids nitidine 7a and fagarone

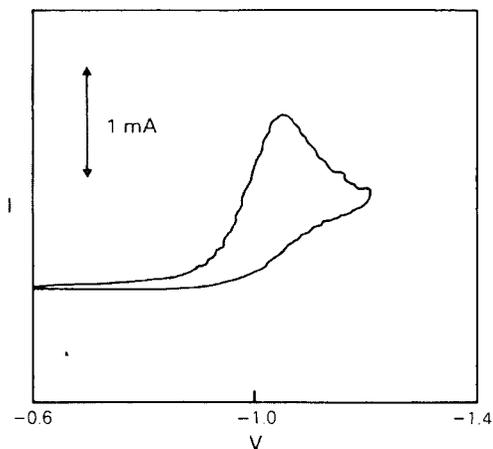


Figure 2 Cyclic voltammogram of 7a in DMF, Pt electrode, scan rate 100 mV/s.

7b, the indenoisoquinolinium salt **8** and 3-nitrobenzothiazolo(3,2-a)quinolinium salts **9a, b**. Studies on **7** and the analogue **8** gave values ranging from -0.90 to -1.15 V (Table III) for **7** and -1.25 to -1.35 V for **8**. The methoxyl substituent is known to result in more negative potentials (Zuman, 1967a). Cyclic voltammetry (CV) (Figure 2) gives irreversible reductions. On the other hand, calculations from polarography (P) and CV indicate reversible behavior. The $E_p - E_{p/2}$ (CV) and $E_{3/4} - E_{1/4}$ (P) values of 60 mV are in reasonable agreement with the theoretical values of 57(CV) and 56(P) mV for a one-electron process. Isoquinolinium salts are known to undergo one electron reduction with formation of the 1,1'-dimer (Bradsher, 1981). The nitrobenzothiazoloquinolinium salts (**9a, b**) give multiple reduction values (Table III) (Figure 3); the most positive range from -0.39 to -0.65 V (irreversible). The more negative waves, about -1.2 V, are reversible. Calculations on the first wave provide values similar to those from **7**, namely, 63(CV) and 60(P) mV. Apparently the reductions are followed by a fast follow-up step. There are two predominant electroactive sites associated with **9**, namely, the nitro group and the iminium ion. The literature $E_{1/2}$ for nitrobenzene is -0.62 V (Wheeler, 1963). Enhancement in the positive direction in our case is due to a more extended, electrophilic system of conjugation. Substitution of methoxyl for hydrogen in the benzothiazole ring has the effect of making the reductions more negative by about 0.03 V as a result of electron donation, in agreement with the reported effect of the 4-methoxyl group in

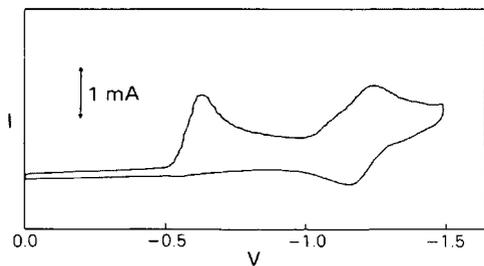


Figure 3 Cyclic voltammogram of **9a** in DMF, Pt electrode, scan rate 100 mV/s.

the 1-phenylpyridinium ion, i.e., $\Delta E_{1/2}$ was more negative by about 0.03 V (Zuman, 1967a).

The activity of **7** has been correlated with the presence of the iminium site (Caolo & Stermitz, 1979), in keeping with our theoretical framework. Also N-methylphenanthridinium salts are known to undergo charge transfer (Parkanyi & Leu, 1975). It is reasonable to associate the activity of **9** in part with nitro or the nitroso reduction product, since compounds of this type are used in cancer therapy (Docampo & Moreno, 1984; Murray & Meyn, 1985). These substances (**7-9**) may exert their activity by binding to DNA (Baez *et al.*, 1983; Cushman *et al.*, 1984). Related anti-tumor alkaloids include coralyne (Cox *et al.*, 1982) and sanguinarine (Nandi & Maiti, 1985).

3. Ellipticines. Most members of this class are anti-tumor agents. Metabolites and various derivatives incorporate quinone-imine and iminium, e.g., **10**. The results from extensive studies (Paoletti *et al.*, 1983) are summarized in Table VII. Electrochemical data demonstrate the ability of the hydroxylated metabolite to function as a charge transfer entity (Paoletti *et al.*, 1983).

Recent reviews deal with iminium ions in the alkaloid category (Knabe, 1979) and from oxidative metabolism of xenobiotics (Overton *et al.*, 1985). The iminium charge transfer theory appears broadly applicable to a wide variety of biologically active agents (Kovacic, 1984), carcinogens (Kovacic *et al.*, in press), drugs (quinoxaline-di-N-oxides) (Ryan *et al.*, 1985), MPTP (Ames *et al.*, in press a), phencyclidine, nicotine and spermine metabolites (Ames *et al.*, in press b), antimalarials (Ames *et al.*, 1985c), mesoionic betaines (Ames *et al.*, 1986d) and benzodiazepines (Crawford *et al.*, in press).

Metal complexes

Metal species are known to elicit a variety of physiological responses. Specific chemical reactions that have been observed include oxygen radical formation (Ames, 1983; Stern, 1985) and DNA strand cleavage (Furst

& Radding, 1984) (Table VII). Formation of complexes with DNA is reported for some cases (Furst & Radding, 1984; Saenger, 1984) (Table VII).

1. *cis*-DDP. The most prominent member of the anti-cancer group is *cis*-DDP. Several reviews summarize much of the work (Roberts & Thomson, 1979; Rosenberg, 1980; Barton & Lippard, 1980). Binding of Pt(II) to guanine of DNA is known to occur, and is believed to have marked biochemical and pharmacological significance (Pinto & Lippard, 1985; Macquet & Theophanides, 1975; Ciccarelli *et al.*, 1985). Considerable effort has been devoted to structural analysis of the DNA-Pt(II) complex (Sherman *et al.*, 1985; Marcellis *et al.*, 1980; Rosenberg, 1980).

Since there is apparent conversion to the diaquodiammine metabolite *in vivo* (Carsey & Boudreaux, 1980), attention was centered on this form in the electroreduction studies. Guanosine was used as the model ligand. Cyclic voltammetry data for the Pt complexes are presented in Table IV. All reductions are irreversible. No reduction occurs before background for *cis*-DDP and guanosine. Experiments with the *cis*-diaquodiammine Pt(II)-guanosine complex (solid or in solution) revealed approximately

the same E_p values (-0.96 to -1.0 V) as for the Pt(II) precursor. The similar results may be due to involvement of supporting electrolyte since added salt is known to alter the 1:1 Pt(II)(H₂O)₂(NH₃)₂-guanoside complex in solution (Marcellis *et al.*, 1980). No reduction occurs before background for *trans*-DDP, and the corresponding diaquo derivative is reduced at -1.20 V. The *trans*-diaquodiammine Pt(II)-guanosine complex (1:1 and 1:2 in solution) gave E_p values that are more negative (-1.3 to -1.6 V). Thus, the reduction potentials in the *cis* series are more positive than for the *trans* counterparts. Since *trans*-DDP is less active (Cleare, 1974) than the *cis*-isomer a correlation exists between potency and ease of electroreduction, in accord with the general mechanistic theme. Prior rationale for the difference in activity has been summarized (Johnson *et al.*, 1985).

The proposed pathway entailing catalytic production of oxy radicals is consistent with effectiveness of the Pt drug at low doses (Rosenberg, 1980; Barton & Lippard, 1980). The toxicity is reduced by mercapto-containing compounds that are well known antioxidants (Nagy *et al.*, 1986; Kempf *et al.*, 1986). Other radical scavengers such as α -tocopherol and *N,N'*-diphenyl-*p*-phenylenediamine exerted a similar effect (Sugihara & Gemba, 1986). The investigators proposed free radical damage by the drug. Also thiols protected against mutagenesis (Nagy *et al.*, 1986) a condition generally attributed to oxy radicals (Kovacic, 1984). There is evidence for a close relationship between mutagenesis and carcinogenesis (Slaga, 1983). Chromosomal aberrations, primarily chromatid breaks, are known to be induced by *cis*-DDP (Flessel *et al.*, 1980).

Table IV Cyclic voltammetry of Pt(II)-guanosine complexes.^a

Compound	$-E_p$ (V)
<i>cis</i> -DDP	NR ^{b,c}
<i>trans</i> -DDP	NR ^{b,c}
Guanosine	NR ^{b,c}
<i>cis</i> -Pt(II)(H ₂ O) ₂ (NH ₃) ₂	0.96
<i>cis</i> -Pt(II)(H ₂ O) ₂ (NH ₃) ₂ - guanosine	0.96
<i>cis</i> -Pt(II)(H ₂ O) ₂ (NH ₃) ₂ - guanosine (1:1 solution)	1.0
<i>trans</i> -Pt(II)(H ₂ O) ₂ (NH ₃) ₂	1.20
<i>trans</i> -Pt(II)(H ₂ O) ₂ (NH ₃) ₂ - guanosine (1:1 solution)	1.45
(1:2 solution)	1.30, 1.60

^a Pt flag, tetraethylammonium perchlorate (0.1 M), substrate (0.5 mM), vs. Ag/AgCl, 100 mV/s.

^b No reduction.

^c 200 mV/s.

2. Complexes of copper and iron

(a) *Copper*: Some copper complexes in this category **11** incorporate salicylaldoximes as chelating agents (Lumme *et al.*, 1984). Reduction potentials for **11a** and **11b** ranged from -0.86 to -0.96 V for the most positive values with Pt as the working electrode, and from -0.71 to -0.86 V with Hg. All of the reductions were irreversible (Table V) (Figure 4).

Table V Cyclic voltammetry of copper(II) salicylaldoxime complexes.^a

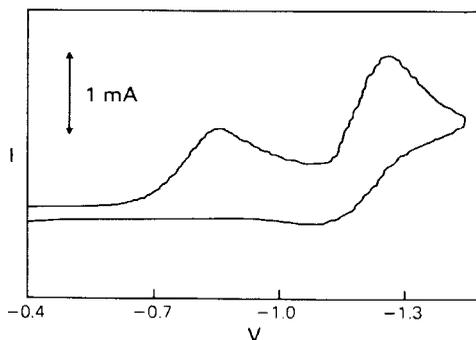
Compound	$-E_p$		Electrode
	DMF	DMSO	
11a	0.86, 1.29	0.93, 1.34	Pt
	0.75, 1.23	0.71, 1.18	Hg
11b	0.91, 1.26	0.97, 1.38	Pt
	0.86 ^b	0.84 ^b , 1.28	Hg

^a 100 mV/s, tetraethylammonium perchlorate (0.1 M), substrate (0.5 mM), irreversible, vs. SCE.

^b Reduction with adsorption.

The difference in reduction potential for **11a** and **11b** (-0.04 to -0.05 V, Pt electrode) is in agreement with the reported effect of the hydroxyl group in anthrone, i.e., $\Delta E_{1/2}$ was more negative by 0.01 to 0.05 V (Zuman, 1967b). There has been a prior suggestion that electron transfer may play a mechanistic role *in vivo* (Lumme & Elo, 1985). For the related Cu(II)(3,5-diisopropylsalicylate)₂, evidence was provided to support the contention that hydrogen peroxide is partly involved in the anti-tumor action (Oberley *et al.*, 1983).

Another class of copper(II) coordination compounds, the thiosemicarbazones, is known to possess anticancer activity (Petering, 1980; Scovill *et al.*, 1982). Reduction of the complex derived from 2-acetylpyridine thiosemicarbazone occurs reversibly at about -0.5 V (Ames *et al.*, 1985c). The

**Figure 4** Cyclic voltammogram of **11a**, in DMF, Pt electrode, scan rate 100 mV/s.

related bis(thiosemicarbazone) complexes display $E_{1/2}$ values of -0.34 to -0.53 V, adjusted to SCE that are attributed to the reduction of Cu(II) to Cu(I) (Winkelman *et al.*, 1974). According to our guiding theme, there is CT resulting in toxic oxy radicals via superoxide. Experimental support is provided by the observation that Cu(I)bis(thiosemicarbazone) is autooxidizable by oxygen (Petering, 1972), a process expected to produce superoxide.

It is relevant that interaction of heterocyclic carboxaldehyde thiosemicarbazones with DNA was observed to result in single strand cleavage (Tsiftoglou *et al.*, 1975); preliminary association of the drug with metal may well occur. DNA scission is commonly associated with oxy radical formation (Demopoulos *et al.*, 1980). Agrawal & Sartorelli (1978) proposed that the action on DNA is of major significance for cytotoxicity.

- (b) *Iron*: Iron complexes of thiosemicarbazones show antineoplastic activity (Scovill *et al.*, 1982). Compound **12** exhibits a reduction wave at -0.23 V (reversible) (Ames *et al.*, 1985c).

Proposals have been made that several well-known agents function after initial coordination with metal ion. The action of bleomycin is summarized in Table VII (Halliwell & Gutteridge, 1985a). According to current thinking (Hecht, 1979; Lown, 1982), the drug sequesters Fe(III) in the cell nucleus and intercalates or binds to DNA. Redox reactions involving the iron and oxygen take place. The reduction potential for the Fe(III) complex is -0.11 V adjusted to SCE (Melnyk *et al.*, 1981).

Adriamycin is known to be a chelating agent for a number of metal ions including Fe(II), Fe(III) and Cu(II) (Halliwell & Gutteridge, 1985a). The iron complexes bind to DNA (Gianni *et al.*, 1985) and reduce molecular oxygen to reactive radicals. DNA cleavage is observed.

3. *Others*. Various other metals, e.g. Rh,

Ru, Sn, Ti, V and Mo, in derivative form exhibit anti-cancer activity (Cleare, 1974; Cleare & Hydes, 1980; Sadler, 1982). However, compared to cis-DDP, they have received relatively little attention. In addition, several agents, such as, α, α' -dipyridyl (Hellman *et al.*, 1983) and picolinic acid (Leuthauser *et al.*, 1982), which are effective against neoplasms, may fit into this mechanistic category based on their ability to bind metals strongly.

Quinones and iminoquinones

Quinone antibiotics have found widespread application in recent years in the treatment of malignancy (Mason, 1982; Lown, 1982; 1983; Waring, 1981). Results from extensive studies, which principally involved anthracyclines, mitomycins, streptonigrin, and saframycins, are summarized in Table VII. The toxicity, found to be oxygen dependent (Halliwell & Gutteridge, 1985a), apparently results from redox cycling of the quinone. Initial metabolic reduction to the semiquinone intermediate, which can bind to DNA (Sinha & Chignell, 1979) evidently is an essential step (Lown, 1982; Emanuel *et al.*, 1984). The overall process has been designated 'site-specific free-radical' generation (Bachur *et al.*, 1982). Inhibition of the rate of DNA scission was observed with added catalase, superoxide dismutase and free radical scavengers (Lown, 1982). However, adriamycin bound to DNA is unable to participate in redox reactions

(Youngman *et al.*, 1984). Strand scission can occur in the absence of binding.

Iminoquinones have not been as extensively studied. Representative members are 5-iminodaunorubicin (Lown *et al.*, 1982), anthrapyrazoles (Fry *et al.*, 1985), and actinomycin D (Halliwell & Gutteridge, 1985a; Doroshov, 1983). Relevant characteristics are intercalation, oxy radical formation, DNA cleavage, and oxygen dependency. As in the quinone case, charge transfer has not been observed after intercalation (Emanuel *et al.*, 1984; Sengupta *et al.*, 1985). Evidence shows that redox cycling and radical generation are less facile with the imine analogues (Lown *et al.*, 1979). Several other anti-cancer agents, e.g., rhodamine 123 (Lampidis *et al.*, 1983), and an oxidative metabolite of ellipticine (Paoletti *et al.*, 1983), possess similar structures.

Table VI contains the reduction potentials for a number of substances in this general category. The $E_{1/2}$ values fall in the range, -0.20 to -1.09 V. A study revealed that the anti-tumor activity of 75% of the investigated iminobenzoquinones could be correctly classified based only on their reduction potentials (Hodnett *et al.*, 1978). Also, the iminoquinones, which exhibit more negative reduction potentials than the quinones, were found to induce less DNA strand cleavage (Lown *et al.*, 1982). The end product of anthrapyrazole reduction is the corresponding dihydro form (Showalter *et al.*, 1986).

Table VI Reduction potentials for some physiologically active quinones and iminoquinones.

Compound	Reduction potential (V)	Reference
Daunorubicin	-0.62	Rao <i>et al.</i> , 1978
Adriamycin	-0.62	Rao <i>et al.</i> , 1978
Mitomycin B	-0.20	Rao <i>et al.</i> , 1977a
Mitomycin C	-0.37	Rao <i>et al.</i> , 1977b
5-Imino-		
daunorubicin	-0.70	Lown <i>et al.</i> , 1982
Anthrapyrazoles	-0.98 to -1.09	Showalter <i>et al.</i> , 1986
Actinomycin D	-0.82	Nakazawa <i>et al.</i> , 1985

Table VII Characteristics of anti-cancer agents.^a

Agent	Generation of reactive oxygen species	DNA binding ^b	DNA cleavage
Quinones	+	+	+
Metals	+	+	+
Bleomycin	+	+	+
Ionizing radiation	+		+
6-Mercaptopurine		+ ^c	+
Alkylating agents	+	+	+
Ellipticine ^d	+	+	+

^a See the discussion for references.^b Intercalation or covalent.^c Nucleotide insertion.^d And derivatives.

The exact state in which these compounds generate oxy radicals at the active site is not established with certainty. Alternatively, the ultimate agent may be a metal complex (*vide supra*).

Radiation

Relevant biological effects are summarized in Table VII (Harman, 1962; Henriksen *et al.*, 1976; Greenstock & Whitehouse, 1984). It is conceivable that indirect generation of oxy radicals also occurs. The nucleic acid bases are considerably more sensitive than the phosphate backbone to radiation (Greenstock & Whitehouse, 1984). Purines are known to form N-oxy species readily on exposure to peroxide (Robins, 1967), which might then serve as CT precursors for radicals (Kovacic *et al.*, in press). For example, adenine 1-oxide displays an $E_{1/2}$ of -0.81 V, pH 1 (Dryhurst, 1977). Also, some forms of ionizing radiation apparently give rise to cationic species that alkylate cellular constituents (Seifter, *et al.*, 1984).

Correlation of reduction potential with physiological activity is not new. Examples include anti-cancer agents (Murray & Meyn, 1985) and other categories (Hodnett *et al.*, 1978; Bogatskii *et al.*, 1971).

There are indications from prior reports that reduction potential *in vivo* may well be more favorable than *in vitro* (Kaye & Stonehill, 1952; Neta *et al.*, 1985). Both dioxidine ($E_{1/2} = -1.06$ V) (Ryan *et al.*, 1985) and

1-methyl-4-phenylpyridinium ion (cyperquat, MPTP metabolite) $E_{1/2} = -1.09$ V) (Ames *et al.*, in press, a) which display rather negative values are reported to function by oxy radical generation via CT (Ryan *et al.*, 1985; (MPTP) Markey *et al.*, 1985). Reversibility is more likely *in vivo* due to immobilization of the CT agent at the active site.

Other considerations

1. Role of oxygen. In our prior discussion, much evidence has been cited for the formation and involvement of activated oxygen species. It is generally believed that superoxide serves as a precursor. Support for this standpoint is provided by investigations on the beneficial influence of oxygen on drug and radiation effectiveness against cancer cells (Cadenas, 1985; Teicher *et al.*, 1981; Gupta & Krishan, 1982). The conclusion was drawn that a common mechanistic pathway pertains for the diverse agents (Gupta & Krishan, 1982; Scheulen & Kappus, 1984) in accord with the present thesis. Drug activity observed during hypoxia (Teicher *et al.*, 1981) can be rationalized by reductive stress involving radical processes (Jones, 1985).

Free radicals derived from oxygen are increasingly implicated in the initiation and progression of various diseases, and in the toxic action of numerous drugs and chemicals (Nelson, 1982; Holtzman, 1982; Sies,

1985; Halliwell & Gutteridge, 1985). The following statement also reflects a unified approach: 'Several of the chemotherapeutic agents are thought to have both their therapeutic and toxic effects by causing an oxidative stress' (Holtzman, 1982). The natural phagocytic response to foreign bodies entails attack by activated oxygen entities (Baehner *et al.*, 1982).

2. *Crucial differences between malignant and normal cells.* As pointed out in the introduction, an important feature of the carcinogen-anti-cancer theory is the cancer-cell property of enhanced susceptibility to reactive oxygen-containing entities. This postulate, advanced quite some time ago, was based primarily on decreased levels of catalase. Since then, other enzymes which destroy these oxy species have been discovered and investigated (Willson, 1983). The superoxide dismutase (SOD) enzyme decomposes superoxide which is generated by aerobic metabolic reactions. Presumably, protection is thereby provided from the adverse effects of oxy radicals, such as hydroxyl, which can arise from the radical anion. In fact, various reports reveal inhibition of radiation carcinogenesis by SOD (Hall & Borek, 1983). A considerable number of studies have found decreased levels of SOD in malignant neoplastic tissues (Oberley & Buettner, 1979). Mn SOD was lower in all cases *vs.* normal cells. The Cu-Zn SOD levels were diminished in many, but not all, tumors. Glutathione peroxidase has also been the object of attention. The basic premise (Kovacic, 1959) advanced more than 26 years ago has been confirmed (Alexander, 1983) and restated after the discovery of the protective role of SOD: 'If equal amounts of superoxide can be delivered to both cancer cells and normal cells, then the cancer cell should be preferentially killed because it has lower Mn SOD activity. Indeed, there is evidence that many of the existing cancer treatments actually are using this rationale because many of the anti-cancer drugs have been shown to produce superoxide' (Oberley & Buettner, 1979). However, other investigators have failed to observe any

obvious relationship between resistance to ionizing radiation or radical-producing drugs and tumor cell content of the following enzymes: Cu-Zn SOD, Mn SOD, catalase, and glutathione peroxidase (Marklund *et al.*, 1982). These findings of large variations in the effectiveness of protective systems may partly account for the observed differences in response by cancer patients.

Another feature of importance is the rate of production of superoxide by tumors. If the generation is similar to or greater than the case of normal cells, then the lowered levels of protective enzymes in the neoplasms would result in enhanced sensitivity to the additional oxidative stress. Investigators have shown that tumor cell mitochondria do produce superoxide (Oberley & Buettner, 1979). In one case, the rate of formation was nearly the same as for normal tissue, whereas in another report there was a five-fold increase.

Although chemotherapy and ionizing radiation have proved beneficial in the treatment of cancer, relapse and limited applicability are commonly seen (Rosenberg, 1980). There are a number of possible rationalizations (Kovacic, 1959). In the context of the theoretical interpretation, increased concentrations of oxy radicals may not be completely effective due to the survival of a small fraction of resistant cancer cells which then proliferate. This is reminiscent of the scenario which has been encountered repeatedly with drugs, insecticides, and herbicides. Furthermore, a fine balance would pertain since the radicals which are generated to combat malignancy are also capable of inducing the same condition. Several recent studies are in harmony with the dual role concept. For instance, the incidence of second cancers in an individual was increased after treatment of the primary ones with anti-cancer drugs (Huang *et al.*, 1983). The induction of new neoplasms was observed as a delayed effect (Harris, 1979). By the same token, initiation of cancer should entail a certain degree of simultaneous inhibition. In fact, early investigators have reported precisely this type of refractory condition on application of carcinogens (Kovacic, 1959).

3. *Alternate mechanisms.* Although the oxy radical theory possesses many attractive features, clearly it presents an oversimplified picture of a complex phenomenon. A number of investigations reveal the important involvement of other factors, principally immunological reactions, inhibition of DNA synthesis, antimetabolite action, and DNA defect repair (Rosenberg, 1980; Roberts & Thomson, 1979; Halliwell & Gutteridge, 1985a; Lumme *et al.*, 1984; Paoletti *et al.*, 1983; Cushman *et al.*, 1984; Baez *et al.*, 1983; Doerr *et al.*, 1961; Remy *et al.*, 1984; Ciccarelli *et al.*, 1985). Specific examples of compounds that are generally believed to operate by other routes are methotrexate (antifolate) (Cole, 1970), α -difluoromethylornithine (DFMO, ornithine decarboxylase inhibitor) (Metcalf *et al.*, 1978) and 5-fluorouracil (antipyrimidine) (Cole, 1970). It is noteworthy that evidence suggests the possibility of CT in some cases. For instance, conjugated iminium species derived from pyridoxal phosphate have been designated as intermediates in the reaction of DFMO with the enzyme (Metcalf *et al.*, 1978). From X-ray data on the binary complex, N-1 protonation of the pteridine portion of methotrexate to iminium is invoked (Bolin *et al.*, 1982). From a study of the ternary complex, the drug and NADPH were shown to be in close proximity (Matthews *et al.* 1978). NADPH might be oxidized by various routes including radical or CT mechanisms (Filman *et al.*, 1982). A metal complex may also participate (Kovacic, 1984) in the case of the pyridoxal imine from DFMO. It is conceivable that several mechanisms operate in concert for certain agents. A recent unifying approach for antineoplastic agents entailed modification of DNA (Hemminki and Ludlum, 1984).

4. *Other biological activity.* In addition to the anti-tumor property, the various agents can display other physiological activities; carcinogenic, mutagenic, cytotoxic, and teratogenic (Magee, 1982; Johnson *et al.*, 1980; Miller & Miller, 1983; Furst & Radding, 1984; Fry, 1983). There is a relationship between antineoplastic activity and the ability to function as drugs in other

areas (Kinnamon *et al.*, 1980). Perhaps some of these responses are also due to oxy radical formation via CT.

5. *Role of antioxidants.* In prior sections, the approach entailed treatment of an established tumor. Alternatively, the problem can be attacked via prevention of initiation by decreasing the concentration of oxy intermediates. Anti-cancer agents in this category, which act as inhibitors of carcinogenesis, would generally be labeled as antioxidants (Demopoulos *et al.*, 1980; Ts'o *et al.*, 1977). A good deal of the work has involved phenolic types, such as butylated hydroxyanisole, selenium compounds, vitamin E, vitamin C, and ethoxyquin. These substances are expected to be ineffective against existing neoplasms, and would act only to inhibit the formation of additional ones from normal cells.

Here again, it is essential to bear in mind the element of specificity. To be effective the antioxidant must reach the site at which the harmful radicals are being generated. Various characteristics of the protective agent would come to bear, including hydrophobic and hydrophilic properties. Hence, it is not surprising that many studies reveal beneficial effects of antioxidants, whereas others (Willet *et al.*, 1984) do not.

In conclusion, the theoretical scheme entails several features common to most anti-cancer agents:

1. Binding to DNA by alkylation, complexation (minor groove), intercalation, or incorporation within the chain as a special purine.
2. Presence of a charge transfer entity in the form of an iminium salt, metal complex, quinone, ArNO_2 or ArNO .
3. Formation of toxic oxy radicals via superoxide generated by electron transfer.
4. Attack of vital cellular constituents by oxy radicals resulting in death of the cancer.

The carcinogen—anti-cancer paradox is rationalized on the basis of similar mechanisms

operating in both cases; many tumor cells are more susceptible than normal ones to the toxic effects of oxy radicals. Antioxidants appear to function by destroying harmful oxy species.

Acknowledgement

This work was supported by grants from the Shaw Research Fund, Graduate School, University of Wisconsin-Milwaukee, Lynn Dennison, and GTE. We thank Professors Barbara Wells, David Petering, and James Otvos for helpful discussions, Professor Mark Cushman for the benzophenanthridine samples, and Dr Hollis Showalter for information prior to publication.

References

- ABSHIRE, C.J. & BERLINGUET, L. (1964). Synthesis of some alkylated adenines as potential antimetabolites. *Canadian Journal of Chemistry*, **42**, 1599.
- AGRAWAL, K.C. & SARTORELLI, A.C. (1978). The chemistry and biological activity of α -(N)-heterocyclic carboxaldehyde thiosemicarbazones. *Progress in Medicinal Chemistry*, **15**, 349.
- ALEXANDER, P. (1983). Can antioxidants facilitate cancer induction? Oxidation reactions involved in host-mediated destruction of cancer cells. In *Radio Protectors and Anticarcinogens*. Nygaard & Simic (eds), p. 575. Academic Press: New York.
- AMES, B.N. (1983). Dietary carcinogens and anticarcinogens. *Science*, **221**, 1256.
- AMES, J.R., CASTAGNOLI, N., JR., RYAN, M.D. & KOVACIC, P. Oxidative ionic metabolites of 1-methyl-4-phenyl-1,2,3,6-tetrahydropyridine (MPTP): Correlation of electroreduction with physiological behavior. *Free Radical Research Communications* (in press, a).
- AMES, J.R., BRANDANGE, S., RODRIGUEZ, B., CASTAGNOLI, N., JR., RYAN, M.D. & KOVACIC, P. Cyclic voltammetry with cyclic iminium ions. Implications for charge transfer with biomolecules (metabolites of nicotine, phenacylidine and spermine). *Bioorganic Chemistry* (in press, b).
- AMES, J.R., RYAN, M.D., KLAYMAN, D.L., & KOVACIC, P. (1985). Charge transfer and oxy radicals in antimalarial action. Quinones, dapsone metabolites, metal complexes, iminium ions and peroxides. *Journal of Free Radicals in Biology and Medicine*, **1**, 353.
- AMES, J.R., POTTS, K.T., RYAN, M.D. & KOVACIC, P. (1986). Conjugated and cross-conjugated mesomeric betaines. Correlation of electroreduction with structure and physiological activity. *Life Sciences*, **39**, 1085.
- BACHUR, N.R., GEE, M.V. & FRIEDMAN, R.D. (1982). Nuclear catalyzed antibiotic free radical formation. *Cancer Research*, **42**, 1078.
- BAEHNER, R.L., BOXER, L.A., INGRAHAM, L.M. (1982). Reduced oxygen byproducts and white blood cells. In *Free Radicals in Biology*. Vol. 5. Pryor (ed.), pp. 91-93. Academic Press: New York.
- BAEZ, A., GONZALEZ, F.A., VAZQUEZ, D. & WARING, M.J. (1983). Interaction between a 3-nitrobenzothiazolo(3,2-a) quinolinium antitumor drug and deoxyribonucleic acid. *Biochemical Pharmacology*, **32**, 2089.
- BARTON, J.K. & LIPPARD, S.J. (1980). Heavy metal interactions with nucleic acids. In *Nucleic Acid-Metal Ion Interactions*. Spiro (ed.), p. 31. Wiley-Interscience: New York.
- BOGATSKII, A.V., ANDRONATI, S.A., GUL'TYAI, V.P. *et al.* (1971). 1-4-Benzodiazepines and their derivatives. V. Polarographic reduction and structure of 1,3-dihydro-2H-1,4-benzodiazepin-2-ones and -benzodiazepine-2-thiones. *Journal of General Chemistry of the USSR*, **41**, 1364.
- BOLIN, J.T., FILMAN, D.J., MATTHEWS, D.A., HAMLIN, R.C. & KRAUT, J. (1982). Crystal structures of *Escherichia coli* and *Lactobacillus casei* dihydrofolate reductase refined at 1.7 Å resolution. *Journal of Biological Chemistry*, **257**, 13650.
- BRADSHAW, C.K. (1981). Quaternary isoquinolinium salts. In *Isoquinolines*. Grethe (ed.), pp. 417-418. Wiley: New York.
- BRUES, A.M. & GUZMAN BARRON, E.S. (1951). Biochemistry of cancer. *Annual Reviews of Biochemistry*, **20**, 343.
- CADENAS, E. (1985). Oxidative stress and formation of excited species. In *Oxidative Stress*. Sies (ed.), p. 318. Academic Press: New York.
- CAOLO, M.A. & STERMITZ, F.R. (1979). Benzophenanthridinium salt equilibria. *Heterocycles*, **12**, 11.
- CARSEY, T.P. & BOUDREAUX, E.A. (1980). The electronic structure of platinum-guanine complexes. *Chemico-Biological Interactions*, **30**, 189.
- CHABNER, B.A. (1981). Antineoplastic agents. Nucleoside analogs. In *Cancer and Chemotherapy, Antineoplastic Agents*. Vol. III. Crooke & Prestayko (eds), p. 3. Academic Press: New York.
- CHRISTIE, N.T., DRAKE, S., MEYN, R.E. &

- NELSON, J.A. (1984). 6-Thioguanine-induced DNA damage as a determinant of cytotoxicity in cultured chinese hamster ovary cells. *Cancer Research*, **44**, 3665.
- CICCARELLI, R.B., SOLOMON, M.J., VARSHAVSKY, A. & LIPPARD, S.J. (1985). *In vivo* effects of cis- and trans-diamminedichloroplatinum(II) on SV 40 chromosomes: differential repair, DNA-protein cross-linking, and inhibition of replication. *Biochemistry*, **24**, 7533.
- CLEARE, M.J. (1974). Transition metal complexes in cancer chemotherapy. *Coordination Chemistry Reviews*, **12**, 349.
- CLEARE, M.J. & HYDES, P.C. (1980). Antitumor properties of metal complexes. In *Metal Ions in Biological Systems*. Vol. 11. Sigel (ed.), p. 1. Marcel Dekker: New York.
- COLE, W.H. (1970). In *Chemotherapy of Cancer*. Lea & Febiger: Philadelphia.
- COX, O., JACKSON, H., VARGAS, V.A. *et al.* (1982). Synthesis and biological activity of benzothiazolo- and benzoxazolo[3,2-a]quinolinium salts. *Journal of Medicinal Chemistry*, **25**, 1378.
- COX, O., JACKSON, H., RIVERA, L.A., RAMIREZ, L., unpublished work.
- CRAWFORD, P.W., KOVACIC, P., GILMAN, N.W. & RYAN, M.D. Charge transfer mechanism for benzodiazepine (BZ) action. Correlation of reduction potential of BZ iminium with structure and drug activity. *Bioelectrochemistry and Bioenergetics* (in press).
- CUSHMAN, M., MOHAN, P. & SMITH, E.C.R. (1984). Synthesis and biological activity of structural analogues of the anticancer benzophenanthridine alkaloid nitidine chloride. *Journal of Medicinal Chemistry*, **27**, 544.
- DEHAND, J. & JORDANOV, J. (1976). Interaction of cis-diamminotolueneplatinum(II) with nucleosides: evidence for guanosine O(6).N(7) chelation by platinum. *Chemical Communications*, 598.
- DEMOPOULOS, H.B., PIETRONIGRO, D.D., FLAMM, E.S. & SELIGMAN, M.L. (1980). The possible role of free radical reactions in carcinogenesis. *Journal of Environmental Pathology and Toxicology*, **3**, 273.
- DOCAMPO, R. & MORENO, S.N.J. (1984). Free radical intermediates in the antiparasitic action of drugs and phagocytic cells. In *Free Radicals in Biology*. Vol. 6. Pryor (ed.), p. 264. Academic Press: Orlando.
- DOERR, I.L., WEMPEN, I., CLARKE, D.A. & FOX, J.J. (1961). Thiation of nucleosides. III. Oxidation of 6-mercaptopyrimidines. *Journal of Organic Chemistry*, **26**, 3401.
- DOROSHOW, J.H. (1983). Effect of anthracycline antibiotics on oxygen radical formation in rat heart. *Cancer Research*, **43**, 460.
- DRYHURST, G. (1969). Electrochemical reduction of 6-thiopurine and related compounds: polarography, voltammetry, and macroscale electrolysis. *Journal of the Electrochemical Society*, **116**, 1357 (and references therein).
- DRYHURST, G. (1977). Electrochemistry of purine derivatives. In *Electrochemistry of Biological Molecules*. Chap. 3. Academic Press: New York.
- EMANUEL, N.M., BOGDANOV, G.N. & ORLOV, V.S. (1984). Free-radical mechanisms in the cytotoxic action of antitumor antibiotics. *Russian Chemical Reviews*, **53**, 1121.
- FILMAN, D.J., BOLIN, J.T., MATTHEWS, D.A., & KRAUT, J. (1982). Crystal structures of *Escherichia coli* and *Lactobacillus casei* dihydrofolate reductase refined at 1.7 Å resolution. *Journal of Biological Chemistry*, **257**, 13663.
- FLESSEL, C.P., FURST, A. & RADDING, S.B. (1980). A comparison of carcinogenic metals. In *Metal Ions in Biological Systems*. Vol. 10. Carcinogenicity and Metal Ions. Sigel (ed.), Chap. 2. Marcel Dekker: New York.
- FLOYD, R.A. (ed.) (1982). *Free Radicals and Cancer*. Marcel Dekker: New York.
- FRIDOVICH, I. (1983). Superoxide radical: an endogenous toxicant. *Reviews of Pharmacology and Toxicology*, **23**, 239.
- FRY, D.W., BORITZKI, T.J., BESSERER, J.A. & JACKSON, R.C. (1985). *In vitro* DNA strand scission and inhibition of nucleic acid synthesis in L1210 leukemia cells by a new class of DNA complexers, the anthra[1,9-cd] pyrazolo-6(2H)-ones (anthrapyrazoles). *Biochemical Pharmacology*, **34**, 3499.
- FRY, R.J.M. (1983). Radiation carcinogenesis. In *Radioprotectors and Anticarcinogens*. Nygaard & Simic (eds), p. 417. Academic Press: New York.
- FURST, A. & RADDING, S.B. (1984). New developments in the study of metal carcinogenesis. *Journal of Environmental Science and Health*, **C2**, 103.
- GIANNI, L., ZWEIER, J.L., LEVY, A. & MYERS, C.E. (1985). Characterization of the cycle of iron-mediated electron transfer from adriamycin to molecular oxygen. *Journal of Biological Chemistry*, **260**, 6820.
- GREENSTOCK, C.L. & WHITEHOUSE, R.P. (1984). Base damage in irradiated DNA and the effect of oxygen. In *Oxygen Radicals in Chemistry and Biology*. Bors, Saran & Tait (eds), p. 619. deGruyter: New York.
- GUPTA, V. & KRISHAN, A. (1982). Effect of oxygen concentration on the growth and drug

- sensitivity of human melanoma cells in soft-agar clonogenic assay. *Cancer Research*, **42**, 1005.
- HALL, E.J. & BOREK, C. (1983). SOD protection against oncogenic transformation. In *Radio-protectors and Anticarcinogens*. Nygaard & Simic (eds), p. 515. Academic Press: New York.
- HALLIWELL, B. & GUTTERIDGE, J.M.C. (1985). *Free Radicals in Biology and Medicine*. (a), Chap. 8. Clarendon Press: Oxford.
- HARMAN, D. (1956). Aging: a theory based on free radical and radiation chemistry. *Journal of Gerontology*, **11**, 298.
- HARMAN, D. (1962). Role of free radicals in mutation, cancer, aging, and the maintenance of life. *Radiation Research*, **16**, 753.
- HARRIS, C.C. (1979). A delayed complication of cancer therapy—cancer. *Journal of the National Cancer Institute*, **63**, 275.
- HECHT, S.M. (ed.) (1979). In: *Bleomycin: Chemical, Biochemical, and Biological Aspects*. Springer-Verlag: New York.
- HELLMAN, R.M., GLASS, J. & NUNEZ, M.T. (1983). α, α' -Dipyridyl as an antineoplastic agent. In: *Structure and Function of Iron Storage, Transport Proteins, Proceedings of the 6th International Conference*. Urushizaki, Aisen & Listowsky (eds), p. 479. Elsevier: Amsterdam; *Chemical Abstracts* (1984), **101**, 48280.
- HEMINKI, K. & LUDLUM, D.B. (1984). Covalent modification of DNA by antineoplastic agents. *Journal of the National Cancer Institute*, **73**, 1021.
- HENRIKSEN, T., BERGENE, R., HEIBERG, A. & SAGSTUEN, E. (1976). Radical reactions in nucleic acids: crystal systems. In *Free Radicals in Biology*. Vol. 2. Pryor (ed.), p. 258. Academic Press: New York.
- HODNETT, E.M., PRAKASH, G. & AMIRMOAZZAMI, J. (1978). Nitrogen analogues of 1,4-benzoquinones. Activities against the ascitic sarcoma 180 of mice. *Journal of Medicinal Chemistry*, **21**, 11.
- HOLMAN, R.A. (1956). Production of abnormal bacteria. Some possible analogies with formation of tumors. *Lancet*, **ii**, 515.
- HOLTZMAN, J.L. (1982). Role of reactive oxygen and metabolite binding in drug toxicity. *Life Sciences*, **30**, 1.
- HUANG, C.C., HAN, C.S., YUE, X.F. *et al.* (1983). Cytotoxicity and sister chromatid exchanges induced *in vitro* by six anticancer drugs developed in the People's Republic of China. *Journal of the National Cancer Institute*, **71**, 841.
- HYSLOP, R.M. & JARDINE, I. (1981). Metabolism of 6-thiopurines. I. Irreversible binding of a metabolite of 6-thiopurine to mammalian hepatic protein *in vitro*. *Journal of Pharmacology and Experimental Therapeutics*, **218**, 621.
- IIO, H., ASAO, K. & TOKOROYAMA, T. (1985). Synthesis of agelasin B and its analogues. *Chemical Communications*, 774.
- ISHIGURO, K., SCHWARTZ, E.L. & SARTORELLI, A.C. (1984). Characterization of the metabolic forms of 6-thioguanine responsible for cytotoxicity and induction of differentiation of HL-60 acute promyelocytic leukemia cells. *Journal of Cellular Physiology*, **121**, 383.
- JOHNSON, N.P., HOESCHELE, J.D., RAHN, R.O., O'NEILL, J.P. & HSIE, A.W. (1980). Mutagenicity, cytotoxicity, and DNA binding of platinum(II) chloroamines in chinese hamster ovary cells. *Cancer Research*, **40**, 1463.
- JOHNSON, N.P., MAZARD, A.M., ESCALIER, J. & MACQUET, J.P. (1985). Mechanism of the reaction between $\text{cis-[PtCl}_2(\text{NH}_3)_2]$ and DNA *in vitro*. *Journal of the American Chemical Society*, **107**, 6376.
- JONES, J.W. & ROBINS, R.K. (1963). Purine nucleosides. III. Methylation studies of certain naturally occurring purine nucleosides. *Journal of the American Chemical Society*, **85**, 193.
- JONES, D.P. (1985). The role of oxygen concentration in oxidative stress: hypoxic and hyperoxic models. In *Oxidative Stress*. Sies (ed.), p. 189. Academic Press: New York.
- KAYE, R.C. & STONEHILL, H.I. (1952). Polarographic reduction of some natural and artificial hydrogen-carriers in bacterial enzyme systems. *Journal of the Chemical Society*, 3244.
- KEMPF, S.R., IVANKOVIC, S., WIESSLER, M. & SCHMAEHL, D. (1986). Effective prevention of the nephrotoxicity of cisplatin (CDDP) by administration of sodium 2-mercaptoethanesulfonate (MESNA) in rats. *British Journal of Cancer*, **52**, 937; *Chemical Abstracts* (1986), **104**, 61639.
- KINNAMON, K.E., STECK, E.A. & RANE, D.S. (1980). Anticancer agents and antitypanosomiasis activity in mice. *Journal of the National Cancer Institute*, **64**, 391.
- KNABE, J. (1979). Iminium salts in nature. In Iminium Salts in Organic Chemistry Part 2, Bohme & Viehe (eds), p. 733. In *Advances in Organic Chemistry*. Taylor (ed.). Wiley-Interscience: New York.
- KOVACIC, P. (1959). An integrated concept of carcinogenic-anticarcinogenic action. *Ohio Journal of Science*, **59**, 318 (and references therein).
- KOVACIC, P. (1960). A proposed role for hydrogen peroxide in carcinogenesis. *Ohio Journal of Science*, **60**, 283.

- KOVACIC, P. (1984). Does charge transfer by diiminium play a widespread role in living systems? *Kemija u Industriji*, **33**, 473 (and references therein).
- KOVACIC, P., CRAWFORD, P.W., RYAN, M.D. & NELSON, V.C. Charge transfer mechanism for carcinogenesis by alkylating and other agents. *Bioelectrochemistry and Bioenergetics* (in press).
- LAMPIDIS, T.J., BERNAL, S.D., SUMMERHAYES, I.C. & CHEN, L.B. (1983). Selective toxicity of rhodamine 123 in carcinoma cells *in vitro*. *Cancer Research*, **43**, 716.
- LEUTHAUSER, S.W.C., OBERLEY, L.W. & OBERLEY, T.D. (1982). Antitumor activity of picolinic acid in CBA/J mice. *Journal of the National Cancer Institute*, **68**, 123.
- LIJINSKY, W. (1976). Interaction with nucleic acids of carcinogenic and mutagenic N-nitroso compounds. *Progress in Nucleic Acid Research and Molecular Biology*, **17**, 247.
- LOWN, J.W., CHEN, H.-H., PLAMBECK, J.A. & ACTON, E.M. (1979). Diminished superoxide anion generation by reduced 5-imino-daunorubicin relative to daunorubicin and the relationship to cardiotoxicity of the anthracycline antitumor agents. *Biochemical Pharmacology*, **28**, 2563.
- LOWN, J.W. (1982). Newer approaches to the study of the mechanisms of action of antitumor antibiotics. *Accounts of Chemical Research*, **15**, 381.
- LOWN, J.W., CHEN, H.-H., PLAMBECK, J.A. & ACTON, E.M. (1982). Further studies on the generation of reactive oxygen species from activated anthracyclines and the relationship to cytotoxic action and cardiotoxic effects. *Biochemical Pharmacology*, **31**, 575.
- LOWN, J.W. (1983). The mechanism of action of quinone antibiotics. *Molecular and Cellular Biochemistry*, **55**, 17.
- LUMME, P. & KORVOLA, M.-L. (1975). Studies on coordination compounds. VI. Thermogravimetric, differential thermogravimetric, differential thermal analysis and mass spectrometric studies of some cobalt(II), nickel(II), and copper(II) salicylaldoximates, 2-indolecarboxylates and 2-thiophenecarboxylates. *Thermochemica Acta*, **13**, 419.
- LUMME, P., ELO, H. & JANNE, J. (1984). Antitumor activity and metal complexes of the first transition series. Trans-bis(salicylaldoximate) copper(II) and related copper(II) complexes, a novel group of potential antitumor agents. *Inorganica Chimica Acta*, **92**, 241.
- LUMME, P. & ELO, H. (1985). Antitumor activity and metal complexes, a comparison. *Inorganica Chimica Acta*, **107**, L15.
- MACQUET, J.P. & THEOPHANIDES, T. (1975). DNA-platinum interactions *in vitro* with trans- and cis-Pt(NH₃)₂Cl₂. *Bioinorganic Chemistry*, **5**, 59.
- MAGEE, P.N. (1982). Interaction of activated intermediates of chemical carcinogens with cellular DNA and its possible prevention. In *Free Radicals, Lipid Peroxidation, and Cancer*. McBrien & Slater (eds), p. 353. Academic Press: New York.
- MARCELIS, A.T.M., VAN KRALINGEN, C.G. & REEDIJK, J. (1980). The interactions of cis- and trans-diammine platinum compounds with 5'-guanosine monophosphate and 5'-deoxyguanosine monophosphate. A proton nmr investigation. *Journal of Inorganic Biochemistry*, **13**, 213.
- MARKEY, S.P., CASTAGNOLI, N., DAVIS, M. *et al.* (eds) (1985). *Symposium on MPTP. A parkinsonian syndrome producing neurotoxin*. Uniformed Services University of the Health Sciences. Bethesda, Maryland.
- MARKLUND, S.L., WESTMAN, N.G., LUNDGREN, E. & ROOS, G. (1982). Copper- and zinc-containing superoxide dismutase, manganese-containing superoxide dismutase, catalase, and glutathione peroxidase in normal and neoplastic human cell lines and normal cell tissues. *Cancer Research*, **42**, 1955.
- MASON, R.P. (1982). Free radical intermediates in the metabolism of toxic chemicals. In *Free Radicals in Biology*. Vol. V. Pryor (ed.), p. 161. Academic Press: New York.
- MATHEWS, D.A., ALDEN, R.A., BOLIN, J.T. *et al.* (1978). Dihydrofolate reductase from *Lactobacillus casei*. *Journal of Biological Chemistry*, **253**, 6946.
- MELNYK, D.L., HORWITZ, S.B. & PEISACH, J. (1981). Redox potential of iron-bleomycin. *Biochemistry*, **20**, 5327.
- METCALF, B.W., BEY, P., DANZIN, C. *et al.* (1978). Catalytic irreversible inhibition of mammalian ornithine decarboxylase (EC 4.1.1.17) by substrate and product analogues. *Journal of the American Chemical Society*, **100**, 2551.
- MILLER, J.A. & MILLER, E.C. (1983). The metabolic activation and nucleic acid adducts of naturally-occurring carcinogens: recent results with ethyl carbamate and the spice flavors safrole and estragole. *British Journal of Cancer*, **48**, 1.
- MURRAY, D. & MEYN, R.E. (1985). DNA damage in normal and neoplastic mouse tissues after treatment with misonidazole *in vivo*. *Biochemical Pharmacology*, **34**, 3275 (and references therein).
- NAGY, B., DALE, P.J., & GRDINA, D.J. (1986).

- Protection against cis-diamminedichloroplatinum toxicity and mutagenicity in V79 cells by 2[(aminopropyl)amino]ethanethiol. *Cancer Research*, **46**, 1132.
- NAKAZAWA, H., BACHUR, N.R., CHOU, F.T.E., MOSSOBA, M.M. & GUTIERREZ, P.L. (1985). Electrochemical and electron spin resonance studies of actinomycin D and other phenoxazones. *Biophysical Chemistry*, **21**, 137.
- NANDI, R. & MAITI, M. (1985). Binding of sanguinarine to deoxyribonucleic acids of differing base composition. *Biochemical Pharmacology*, **34**, 321.
- NELSON, S.D. (1982). Metabolic activation and drug toxicity. *Journal of Medicinal Chemistry*, **25**, 753.
- NETA, P., RICHOUX, M.-C. & HARRIMAN, A. (1985). Intramolecular association of covalently linked viologen radicals. *Journal of the Chemical Society, Faraday Transactions 2*, **81**, 1427.
- OBERLEY, L.W. & BUETTNER, G.R. (1979). Role of superoxide dismutase in cancer: a review. *Cancer Research*, **39**, 1141.
- OBERLEY, L.W., ROGERS, K.L., SCHUTT, L., OBERLEY, T.D., LEUTHAUSER, S.W.C., & SORENSON, J.R.J. (1983). Possible role of glutathione in the antitumor effect of a copper containing synthetic superoxide dismutase in mice. *Journal of the National Cancer Institute*, **71**, 1089.
- OVERTON, M., HICKMAN, J.A., THREADGILL, M.D. *et al.* (1985). The generation of potentially toxic, reactive iminium ions from the oxidative metabolism of xenobiotic N-alkyl compounds. *Biochemical Pharmacology*, **34**, 2055.
- PAOLETTI, C., AUCLAIR, C. & MEUNIER, B. (1983). Biochemical mechanisms of the cytoxic action of the antitumor ellipticine derivatives. In *Mechanism of Drug Action*. Singer, Mansour & Ondraza (eds), p. 305. Academic Press: New York.
- PARKANYI, C. & LEU, G.J. (1975). Charge-transfer spectra of some heterocyclic cations. *Zeitschrift für Naturforschung*, **30b**, 984.
- PETERING, D.H. (1972). The reaction of 3-ethoxy-2-oxobutyraldehyde bis (thiosemicarbazonato) copper(II) with thiols. *Bioinorganic Chemistry*, **1**, 273.
- PETERING, D.H. (1980). Carcinostatic copper complexes. In *Metal Ions in Biological Systems*. Vol. 11. Sigel (ed.), p. 197. Marcel Dekker: New York.
- PINTO, A.L. & LIPPARD, S.J. (1985). Binding of the antitumor drug cis-diamminedichloroplatinum(II) (cisplatin) to DNA. *Biochimica et Biophysica Acta*, **780**, 167.
- RAJALAKSHMI, S., RAO, P.M. & SARMA, D.S.R. (1982). Chemical carcinogenesis: inter- actions of carcinogens with nucleic acids. In *Cancer* 1. 2nd edn. Becker (ed.), p. 335ff. Plenum Press: New York.
- RAO, G.M., LOWN, J.W. & PLAMBECK, J.A. (1977a). Electrochemical studies of antitumor antibiotics. I. Cyclic voltammetry study of mitomycin B. *Journal of the Electrochemical Society*, **124**, 195.
- RAO, G.M., BEGLEITER, A., LOWN, J.W. & PLAMBECK, J.A. (1977b). Electrochemical studies of antitumor antibiotics. II. Polarographic and cyclic voltammetric studies of mitomycin C. *Journal of the Electrochemical Society*, **124**, 199.
- RAO, G.M., LOWN, J.W. & PLAMBECK, J.A. (1978). Electrochemical studies of antitumor antibiotics. III. Daunorubicin and adriamycin. *Journal of the Electrochemical Society*, **125**, 534.
- REICH, S.D. (1981). Other alkylating agents. In *Cancer and Chemotherapy, Antineoplastic Agents*. Vol. III. Croke & Prestayko (eds), p. 61. Academic Press: New York.
- REMY, J.J., BELEHRADEK, J. & JACQUEMINE-SABLON, A. (1984). Expression of drug sensitivity and tumorigenicity in intra-species hybrids between 9-hydroxyellipticine-sensitive and -resistant cells. *Cancer Research*, **44**, 4587.
- ROBERTS, J.J. & THOMSON, A.J. (1979). The mechanism of action of antitumor platinum compounds. *Progress in Nucleic Acid Research and Molecular Biology*, **22**, 71.
- ROBINS, R.K. (1967). The purines and related ring systems. In *Heterocyclic Compounds*. Vol. 8. Elderfield (ed.), pp. 356-357. Wiley: New York.
- ROSENBERG, B. (1980). Platinum complexes for the treatment of cancer. In *Nucleic Acid-Metal Ion Interactions*. Spiro (ed.), p. 1. Wiley-Interscience: New York.
- RYAN, M.D., SCAMEHORN, R.G. & KOVACIC, P. (1985). Charge transfer in the mechanism of drug action involving quinoxaline di-N-oxides. *Journal of Pharmaceutical Sciences*, **74**, 492.
- SADLER, P.J. (1982). Inorganic pharmacology. *Chemistry in Britain*, **18**, 182.
- SAENGER, W. (1984). *Principles of Nucleic Acid Structure*. p. 201. Springer-Verlag: New York.
- SCHEULEN, M.E. & KAPPUS, H. (1984). The activation of oxygen by bleomycin is catalyzed by NADPH-cytochrome P-450 reductase in the presence of iron ions and NADPH. In *Oxygen Radicals in Chemistry and Biology*. Bors, Saran & Tait (eds), p. 425. deGruyter: New York.
- SCOVILL, J.P., KLAYMAN, D.L. & FRANCHINO, C.F. (1982). 2-Acetylpyridine thiosemicarbazones. 4. Complexes with transi-

- tion metals as antimalarial and antileukemic agents. *Journal of Medicinal Chemistry*, **25**, 1261.
- SEIFTER, E., RETTURA, G., PADAWER, J. *et al.* (1984). Morbidity and mortality reduction by supplemental vitamin A or β -carotene in CBA mice given total-body γ -radiation. *Journal of the National Cancer Institute*, **73**, 1167.
- SENGUPTA, S.K., KELLY, C. & SEHGAL, R.K. (1985). "Reverse" and "symmetrical" analogs of actinomycin D: metabolic activation and *in vitro* and *in vivo* tumor growth inhibitory activities. *Journal of Medicinal Chemistry*, **28**, 620.
- SHERMAN, S.E., GIBSON, D., WANG, A.H.-J. & LIPPARD, S.J. (1985). X-ray structure of the major adduct of the anticancer drug cisplatin with DNA: cis-[Pt(NH₃)₂{d(pGpG)}]. *Science*, **230**, 412.
- SHOOTER, K.V. (1972). Some aspects of the interaction of carcinogenic and mutagenic agents with purines in nucleic acids. In *The Purines—Theory and Experiment, The Jerusalem Symposia on Quantum Chemistry and Biochemistry*. Vol. IV. Bergmann & Pullman (eds), p. 509. The Israel Academy of Sciences and Humanities: Jerusalem.
- SHOWALTER, H.D.H., FRY, D.W., LEOPOLD, W.R., LOWN, J.W., PLAMBECK, J.A. & RESZKA, K. (1986). Design, biochemical pharmacology, electrochemistry, and tumor biology of antitumor anthrapyrazoles. *Anti-Cancer Drug Design*, **1**, 73.
- SIES, H. (ed.) (1985). In *Oxidative Stress*. Academic Press: New York.
- SINHA, B.K. & CHIGNELL, C.F. (1979). Binding mode of chemically activated semiquinone free radicals from quinone anticancer agents to DNA. *Chemico-Biological Interactions*, **28**, 301.
- SLAGA, T.J. (1983). Chemical carcinogenesis and anticarcinogenesis. In *Radioprotectors and Anticarcinogens*. Nygaard & Simic (eds), p. 437. Academic Press: New York.
- STERN, A. (1985). Red cell oxidative damage. In *Oxidative Stress*. Sies (ed.), pp. 340–341. Academic Press: New York.
- SUGIHARA, K. & GEMBA, M. (1986). Modification of cisplatin toxicity by antioxidants. *Japanese Journal of Pharmacology*, **40**, 353; *Chemical Abstracts* (1986), **104**, 81676.
- TEICHER, B.A., LAZO, J.S. & SARTORELLI, A.C. (1981). Classification of antineoplastic agents by their selective toxicities toward oxygenated and hypoxic tumor cells. *Cancer Research*, **41**, 73.
- TSITSOGLOU, A.S., HWANG, K.M., AGRAWAL, K.C. & SARTORELLI, A.C. (1975). Strand scission of sarcoma 180 tumor cell DNA induced by 1-formylisoquinoline thiosemicarbazone. *Biochemical Pharmacology*, **24**, 1631.
- TS'O, P.O.P., CASPARY, W.J. & LORENTZEN, R.J. (1977). The involvement of free radicals in chemical carcinogenesis. In *Free Radicals in Biology*. Vol. III. Pryor (ed.), p. 294. Academic Press: New York.
- WARBURG, O., GAWEHN, K. & GEISSLER, A.-W. (1957). The effect of hydrogen peroxide on cancer cells and on embryonic cells. *Zeitschrift für Naturforschung*, **12b**, 393.
- WARING, M.J. (1981). DNA modification and cancer. *Annual Reviews of Biochemistry*, **50**, 159.
- WHEELER, O.H. (1963). Polarographic reduction of *o*-alkylnitrobenzenes. *Canadian Journal of Chemistry*, **41**, 192.
- WILLET, W.C., POLK, B.F., UNDERWOOD, B.A. *et al.* (1984). Relation of serum vitamins A and E and carotenoids to the risk of cancer. *The New England Journal of Medicine*, **310**, 430.
- WILLSON, R.L. (1983). Free radical repair mechanisms and the interactions of glutathione and vitamins C and E. In *Radioprotectors and Anticarcinogens*. Nygaard & Simic (eds), p. 1. Academic Press: New York.
- WINKELMANN, D.A., BERMKE, Y. & PETERING, D.H. (1974). Comparative properties of the antineoplastic agent, 3-ethoxy-2-oxobutyraldehyde bis (thiosemicarbazonato) copper(II) and related chelates: linear free energy correlations. *Bioinorganic Chemistry*, **3**, 261.
- YAO, T., WASA, T. & MUSHA, S. (1976). The electrochemical behavior of the purine bases and their nucleosides in *N,N*-dimethylformamide. *Nippon Kagaku Kaishi*, 704; *Chemical Abstracts* (1976), **85**, 11616.
- YOUNGMAN, R.J., GOTZ, F. & ELSTNER, E.F. (1984). Role of oxygen activation in adriamycin-mediated DNA strand scission and the effect of binding on the redox properties of the drug. In *Oxygen Radicals in Chemistry and Biology*. Bors, Saran & Tait (eds), p. 131. deGruyter: Berlin.
- ZUMAN, P. (1967). *Substituent Effects in Organic Polarography*. (a) p. 148, (b) p. 229. Plenum Press: New York.