Chronic Stress and Ovarian Function in Female Childhood Cancer Survivors

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Chronic Stress and Ovarian Function in Female Childhood Cancer Survivors

Theresa M. Hardy, PhD, RN, Mauricio Garnier-Villarreal, PhD, Jennifer M. Ohlendorf, PhD, RN, and Donna O. McCarthy, PhD, RN

OBJECTIVES: To explore the relationships among perceived stress, biomarkers of hypothalamic–pituitary–adrenal (HPA) activity, gonadotropin levels, and anti-Müllerian hormone (AMH) in female childhood cancer survivors (CCSs).

SAMPLE & SETTING: 24 female CCSs from the Royal Hospital for Sick Children in Edinburgh, Scotland, were included in the study.

METHODS & VARIABLES: Perceived stress was measured using the Perceived Stress Scale. HPA activity was measured using salivary cortisol and hair cortisol. Ovarian function was measured using serum gonadotropin levels and serum AMH levels. Latent growth curve modeling was used to determine diurnal cortisol slope and intercept. Bayesian structural equation modeling was used to explore the relationship among perceived stress, biomarkers of HPA activity, and ovarian function.

RESULTS: The authors found an inverse association between perceived stress and ovarian function and a positive association between biomarkers of HPA activity and ovarian function.

IMPLICATIONS FOR NURSING: Further research is needed to understand factors contributing to risk for post-treatment reproductive dysfunction in female CCSs.

KEYWORDS: childhood cancer survivors; ovarian function; biomarkers; perceived stress

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An estimated 1 in 1,000 adults aged younger than 35 years is a survivor of childhood cancer (Blumenfeld, 2012). Because of significant advances in the treatment of pediatric cancer, the five-year survival rate exceeds 80% in the United States (Phillips et al., 2015). With an increasing number of survivors, there has been growing recognition of the late effects of cancer treatment (Kremer et al., 2013). Among these, reproductive dysfunction is a major concern for cancer survivors and is highly correlated with quality of life in this population (Cherven, Mertens, Wasilewski-Masker, Williamson, & Meacham, 2015; Knopman, Papadopoulos, Grifo, Fino, & Noyes, 2010; Kondapalli et al., 2014; Letourneau, Chan, & Rosen, 2013). Among female childhood cancer survivors (CCSs), 6% experience acute ovarian failure, and another 23% experience a significant reduction in ovarian function (Salih et al., 2015).

In clinical practice, post-treatment ovarian function is assessed using a profile of hormones, including follicle-stimulating hormone (FSH) and luteinizing hormone (LH), plus the presence or absence of menses. However, neither FSH nor menstrual cyclicity post–cancer treatment are reliable predictors of future fertility (Knight et al., 2015). Because risk of infertility related to premature ovarian failure is associated with the size of the ovarian follicle pool (ovarian reserve), a biomarker that more closely reflects the number of remaining follicles in the ovary would have significant clinical potential. Anti-Müllerian hormone (AMH) is a biomarker of the ovarian reserve; plasma levels reflect the continuous noncyclic growth of small follicles and, therefore, mirror the size of the remaining follicle pool (Jeppesen et al., 2013). AMH is prematurely reduced in CCSs (Anderson & Wallace, 2013; Charpentier et al., 2014; Miyoshi et al., 2013) and may be an early marker of significant gonadotoxicity post-treatment (Brougham et al., 2012; Lie Fong et al., 2009). Although studies have demonstrated the use of pretreatment...

Cancer survivors have high rates of psychological stress and hypothalamic-pituitary-adrenal (HPA) dysregulation (Oancea et al., 2014; Taylor, Absolom, Snowden, & Eiser, 2012; Zeltzer et al., 2009). One of the primary regulators of reproductive function is the HPA axis. Psychological stress impairs reproductive function by activating the HPA axis, which suppresses hypothalamic pituitary gonadal function (Kalantaridou et al., 2010; Louis et al., 2011; Lynch, Sundaram, Maisog, Sweeney, & Buck Louis, 2014; Whirledge & Cidlowski, 2013). Increased HPA activity is associated with elevated levels of glucocorticoids, such as cortisol. At homeostatic levels, cortisol contributes to steroid biosynthesis and maintenance of gonadotropin release; at elevated levels, it suppresses gonadotropin-releasing hormone secretion, reduces pulsatile LH secretion, and increases rates of follicle atresia (Breen & Mellon, 2014; Whirledge & Cidlowski, 2013). Because higher rates of psychological stress and reproductive dysfunction are observed in cancer survivors, the purpose of this study was to explore relationships among perceived stress, HPA activity, and ovarian function in a sample of female CCSs.

Methods
The sample for this exploratory cross-sectional study consisted of female CCSs aged from 16 to 35 years, who were previously treated for cancer at the Edinburgh Children’s Cancer Centre at the Royal Hospital for Sick Children in Scotland. The inclusion criteria were female patients treated for cancer at the Edinburgh Children’s Cancer Centre who were aged younger than 18 years at cancer diagnosis, were aged from 16 to 35 years at the time of the study visit, were at least one year from completing treatment, and signed written informed consent. The exclusion criteria were a positive pregnancy test, ovarian surgery in the past six months, hormonal therapy in the past three months (GnRH agonists, recFSH), alcohol or drug abuse/dependence according to ICD-10 criteria, or having received an investigational drug in the past three months.

The Marquette University Institutional Review Board and the South East Scotland Research Ethics Committee approved this study. Data management was compliant with the most current guidelines of the Health Insurance Portability and Accountability Act of 1996 and in full conformity with the principles of the Declaration of Helsinki, in accordance with the principles laid down in the Convention of the Council of Europe for the protection of human rights and dignity of the human being. Original patient names and hospital bar codes were deidentified by using a new, independent numbering system for all study-related documentation. A link-list was established and kept separate from other study documentation. The link-list was only accessible to the study investigators and was kept in a locked office at the Royal Hospital for Sick Children.

Instruments
Assessment of ovarian function included serum gonadotropin levels (LH and FSH), estradiol (E2), and AMH. Serum levels of these hormones are routinely measured during the annual follow-up appointment. LH, FSH, and E2 levels are measured to screen for gonadotropin deficiency and ovarian dysfunction post–cancer treatment (Metzger et al., 2013). The laboratory results were retrieved from participants’ medical records. Serum AMH levels were analyzed using the Beckman Coulter AMH Gen II ELISA. The AMH Gen II ELISA has a sensitivity of 0.57 pmol/L.

HPA activity was determined using measures of salivary cortisol and hair cortisol. Salivary cortisol has been used to measure HPA activity in cancer survivors and other populations (Du et al., 2013; Ho, Fong, Chan, & Chan, 2013; Sephton et al., 2013). Cortisol secretory activity follows a diurnal pattern, with levels peaking after awakening and lowest levels before going to bed (Kirschbaum & Hellhammer, 1989). Chronic stress is associated with alterations in the cortisol awakening response (Dedovic & Ngiam, 2015; Révész et al., 2013) and blunting of the morning–evening diurnal slope (Garland, Beck, Lipschitz, & Nakamura, 2015; Ho et al., 2013; Schrepf et al., 2015). A commercially available ELISA kit by Salimetrics® was used for the quantification of cortisol in saliva. This cortisol assay kit has a lower detection limit of 7 ng/dl. The mean intra-assay coefficient is 5.7%, and the mean inter-assay coefficient is 10%.

Chronic stress has also been associated with elevated levels of cortisol in hair (Russell, Koren, Rieder, & Van Uum, 2012; Wells et al., 2014). Hair has a predictable growth rate of 1 cm per month; the most proximal 1 cm segment to the scalp approximates the past month’s cortisol production, the second most proximal segment the month prior to that, and so on (Russell et al., 2012). Hair (3 cm) was processed for extraction of cortisol in the laboratory of
Clemens-Kirschbaum in Dresden, Germany, as previously described (Kirschbaum, Tietze, Skoluda, & Dettenborn, 2009).

Perceived stress was measured using the Perceived Stress Scale (PSS-10) (Cohen, Kamarck, & Mermelstein, 1983). The PSS-10 is a 10-item self-report scale measuring life stress during the past four weeks and is the most widely used measure of this concept. The PSS-10 was designed for use in community samples of participants with education as low as junior high. Good to excellent reliability has been observed with the PSS-10 in varied populations (Cohen et al., 1983), with a Cronbach alpha ranging from 0.78–0.91 (Lee, 2012). The measure has been demonstrated to reliably measure stress appraisal in cancer survivors (Golden-Kreutz, Browne, Frierson, & Andersen, 2009).

Data Collection

Recruitment was conducted by sending letters of invitation to female CCSs in long-term follow-up two weeks prior to their annual review appointment. After providing informed consent, saliva-collecting devices and detailed instructions for collection of samples were provided to participants at their appointment. Participants were instructed not to eat, smoke, consume caffeine, or exercise for 30 minutes prior to sample collection (Schrepf et al., 2015). Participants provided three timed saliva samples for the analysis of salivary cortisol: the first sample was collected before going to bed on the night prior to the study visit, the second immediately upon awakening the morning of their visit before getting out of bed, and the third 30 minutes after the second sample. Participants were instructed to record the time of saliva collection on their saliva-collecting devices and to keep the saliva samples refrigerated until coming in for their study visit to preserve sample integrity. Participants brought the three saliva samples to their study visit. Samples were stored at the Royal Hospital for Sick Children at −20°C. Within a week of collection, a research nurse picked them up and brought them to the Queens Medical Research Institute, where they were centrifuged and stored at −80°C.

During the study visit, participants provided demographic information, including age, ethnicity/race, level of education, marital status, and employment status. The following clinical information was retrieved from participants’ medical records: cancer diagnosis, age at cancer diagnosis, chemotherapy/radiation received, and age at menarche. Participants then completed the PSS-10 (Cohen et al., 1983). Lastly, participants provided a sample of hair for measurement of hair cortisol. A small sample of hair, 20 mg (100–150 strands), was taken from the vertex posterior of the participant’s scalp using a new pair of thinning shears (Hoffman, Karban, Benitez, Goodteacher, & Laudenslager, 2014; Van Uum et al., 2008). Hair length was recorded, and proximal ends of the hair samples were marked. Hair samples were stored in aluminum foil for protection before processing (Hoffman et al., 2014) and labeled with a unique participant study ID.

Data Analysis and Interpretation

Means and standard deviations for all demographic and clinical variables were reported to describe the sample and reveal distributional characteristics. Latent growth curve modeling was used to define diurnal cortisol change over time (three time points). Change processes are determined by combining the underlying pattern of growth (slope) with the initial or baseline measurement (intercept). The intercept and slope were then incorporated into the overall model as independent predictors of ovarian function.

Bayesian structural equation modeling (SEM) was used to explore the relationship between perceived stress and HPA activity and to explore relationships between perceived stress, biomarkers of HPA activity, gonadotropin levels, and AMH (see Figure 1). Bayesian SEM was selected because it facilitates greater precision in estimating the posterior distribution of the parameters for small samples. The posterior distribution describes the best estimate of the parameters.

Note. Covariates consist of age, body mass index, age at menarche, and age at diagnosis.

FIGURE 1. Bayesian Path Analysis Model

![Bayesian Path Analysis Model Diagram]
given the data and the level of uncertainty about the respective parameter by the width of it, represented by the credible interval (Yuan & MacKinnon, 2009). Bayesian SEM also offers the advantage of incorporating previous knowledge about the model’s parameters through the use of priors. The term “prior” is used to refer to the expected parameter distribution based on theory or previous research. Given the exploratory nature of this study, the authors examined the model parameters using weakly informed priors. Weakly informed priors are not intended to guide the parameters, but rather to provide information to delimitate the most likely data space for the parameters. The authors used a special case of SEM called path analysis. In path analysis, every variable in the model is directly measured or observed (Bryan, Schmiege, & Broaddus, 2007). The authors ran path analysis models with salivary cortisol intercept and slope, PSS-10, hair cortisol, body mass index, age, age at diagnosis, and age at menarche as predictors and with AMH, E2, FSH, and LH as outcomes.

Data augmentation was used to account for missing data. Missing data were treated simultaneously as a parameter and data, filling any missing data points with the most likely value given the posterior distribution (Merkle, 2011). Data analysis was conducted in R (Core Team, 2017). Bayesian path analysis was run with the blavaan R package (Merkle & Rosseel, 2018), which estimates the model with the general Bayesian software JAGS (Plummer, 2003). Convergence of the Markov chains was determined using the potential scale reduction factor, also known as univariate R-hat (Gelman & Rubin, 1992). Model convergence was achieved when R-hat was lower than 1.1 for every parameter (Brooks & Gelman, 1998). The model was run with three chains, keeping the last 5,000 iterations from each chain to build the posterior distributions.

**Results**

Twenty-four female CCSs (\( \bar{X} \) age = 21.79 years, SD = 5.68) participated in the study. The majority had at least a high school education (n = 15) and were unmarried (n = 19) (see Table 1). Childhood cancer diagnoses varied considerably and included osteosarcoma (n = 1), rhabdomyosarcoma (n = 5), Wilms tumor (n = 1), optic chiasmal glioma (n = 1), non-Hodgkin lymphoma (n = 3), Ewing sarcoma (n = 2), immature teratoma of the left ovary (n = 1), B-cell lymphoma (n = 1), Hodgkin lymphoma (n = 4), right temporal lobe pilocytic astrocytoma (n = 1), pilocytic astrocytoma of the conus (n = 1), left renal tumor (n = 1), undifferentiated sarcoma of posterior sacral region (n = 1), and acute lymphoblastic leukemia (n = 1). Cancer treatments included chemotherapy only (n = 11), radiation only (n = 1), chemotherapy and radiation (n = 8), or other treatment (n = 4). Mean length of treatment was 0.63 years (SD = 1.05).

**Latent Growth Curve**

The average salivary cortisol intercept (corresponding to the night time point) was 105.25 ng/dl, with a variance of 48.53 ng/dl. On average, salivary cortisol levels increased 24.19 ng/dl per hour with a variance of 9.41 ng/dl. The latent growth curve demonstrated heterogeneity of variance over time (see Figure 2), with increased variability observed at the later time points. Some slopes were positive and others flat, creating an overdispersion at the later time points.

<table>
<thead>
<tr>
<th>TABLE 1. Clinical and Demographic Descriptive Statistics</th>
</tr>
</thead>
<tbody>
<tr>
<td><strong>Variable</strong></td>
</tr>
<tr>
<td>Age (years)</td>
</tr>
<tr>
<td>Age at diagnosis (years)</td>
</tr>
<tr>
<td>Age at menarche (years)</td>
</tr>
<tr>
<td>Body mass index (kg/m²)</td>
</tr>
<tr>
<td>Perceived Stress Scale total</td>
</tr>
<tr>
<td>Night cortisol (ng/dl)</td>
</tr>
<tr>
<td>Morning cortisol (ng/dl)</td>
</tr>
<tr>
<td>Morning cortisol plus 30 minutes (ng/dl)</td>
</tr>
<tr>
<td>Hair cortisol (pg/mg)</td>
</tr>
<tr>
<td>Follicle-stimulating hormone (U/L)</td>
</tr>
<tr>
<td>Luteinizing hormone (U/L)</td>
</tr>
<tr>
<td>Estradiol (pmol/L)</td>
</tr>
<tr>
<td>Anti-Müllerian hormone (pmol/L)</td>
</tr>
</tbody>
</table>
Bayesian Structural Equation Modeling
The authors found that the model predictors (i.e., salivary cortisol intercept, salivary cortisol slope, hair cortisol, and PSS-10) explained 42% of the variance of FSH, 39% of LH, 33% of AMH, and 0% of E2. The standardized regression coefficients allowed the authors to compare the effect of different predictors for each outcome. For AMH, age, age at diagnosis, perceived stress, and hair cortisol levels were the strongest predictors. For FSH, perceived stress was the strongest predictor, followed by the intercept of salivary cortisol and age at menarche. For LH, the authors found that the three predictors related to cortisol (i.e., salivary cortisol intercept, salivary cortisol slope, and hair cortisol) and age at diagnosis were the strongest predictors. For E2, all predictors were equivalent around 0, demonstrating that these predictors did not characterize the behavior of E2.

Discussion
In this small sample of female CCSs, the mean PSS-10 score was 16.43 (SD = 6.73), which is higher than the mean threshold for stress in the general population (13.7 for women) (Cohen et al., 1983). Surviving a life-threatening illness, such as pediatric cancer, represents a major early life stressor (Laufer et al., 2012). Several studies have demonstrated that CCSs report higher levels of stress than healthy controls (Brown, Madan-Swain, & Lambert, 2003; Oancea et al., 2014). Keir, Swartz, and Friedman (2007) demonstrated that long-term cancer survivors are just as likely as patients with cancer to report high levels of stress, suggesting that the time since diagnosis and treatment does not mitigate the psychological stress experienced by cancer survivors.

The authors observed higher mean night salivary cortisol concentrations (greater than 100 ng/dl) and flattened diurnal cortisol slopes in the current sample of female CCSs. In healthy individuals, cortisol concentrations peak in the morning about 30 minutes after awakening and decline throughout the day (Bergen et al., 2012). Higher nighttime cortisol levels and blunted cortisol slopes are abnormal diurnal patterns. It is hypothesized that these abnormal diurnal patterns induce systemic inflammation, which is associated with increased incidence of adverse long-term health outcomes for cancer survivors (e.g., cancer recurrence, chronic symptom experience) (Schrepf et al., 2015).

To the authors’ knowledge, no studies have examined hair cortisol levels in CCSs. In this small sample of 24 CCSs, the authors found a wide range of three-month hair cortisol concentrations (range = 1.08–211.7 pg/mg, \( \bar{X} = 19.94 \) pg/mg, SD = 43.23 pg/mg). There was one outlier in the sample (211.7 pg/mg); excluding the outlier, the range of hair cortisol concentrations was 1.08 pg/mg to 55.71 pg/mg. Age may contribute to changes in hair cortisol concentrations (Gow, Thomson, Rieder, Van Uum, & Koren, 2010); however, no association between age and cortisol concentrations was observed in the current sample. The mean hair cortisol concentration observed in this sample is lower than the mean reported in healthy samples (Stalder & Kirschbaum, 2012). In a meta-analysis of chronic stress and HPA activity, Miller, Chen, and Zhou (2007) reported that patterns of HPA activity in response to chronic stress vary by time since onset. When the eliciting stimulus of a chronic stressor first begins, there is an initial activation of the HPA axis, which results in elevated concentrations of cortisol. As time passes, this activity lessens, and cortisol secretion rebounds to below normal (Miller et al., 2007). This may explain the lower mean hair cortisol concentrations observed in this sample of CCSs. Because hypocortisolism and hypercortisolism are associated with adverse health outcomes, the finding of lower mean hair cortisol concentrations in this sample is notable and requires further investigation (Miller et al., 2007).
In this study of 24 CCSs, the authors found that the model predictors explained 33% of the variation in AMH levels, with the highest standardized regression coefficients observed for age, age at diagnosis, perceived stress, and hair cortisol (see Table 2). AMH was inversely associated with age, age at diagnosis, and perceived stress; with an increase of one standard deviation in age, age at diagnosis, and perceived stress score, AMH declined 5.66 pmol/L, 5.19 pmol/L, and 5.02 pmol/L, respectively. The inverse association between age and AMH is well established in the literature (Kelsey, Wright, Nelson, Anderson, & Wallace, 2011). Barton et al. (2013) also found that older age at diagnosis increased risk of post-treatment infertility.

### TABLE 2. Bayesian Path Analysis by Outcome

<table>
<thead>
<tr>
<th>Predictor</th>
<th>β</th>
<th>95% CI</th>
<th>Standardized Estimate</th>
</tr>
</thead>
<tbody>
<tr>
<td><strong>Anti-Müllerian hormone</strong></td>
<td></td>
<td></td>
<td></td>
</tr>
<tr>
<td>Salivary cortisol intercept</td>
<td>4.138</td>
<td>[-10.39, 18.81]</td>
<td>0.133</td>
</tr>
<tr>
<td>Salivary cortisol slope</td>
<td>5.26</td>
<td>[-7.14, 18.28]</td>
<td>0.082</td>
</tr>
<tr>
<td>Perceived Stress Scale</td>
<td>-5.02</td>
<td>[-10.93, 1.58]</td>
<td>-0.287*</td>
</tr>
<tr>
<td>Hair cortisol</td>
<td>3.757</td>
<td>[-2.43, 10.57]</td>
<td>0.214*</td>
</tr>
<tr>
<td>Body mass index</td>
<td>1.955</td>
<td>[-4.65, 8.29]</td>
<td>0.112</td>
</tr>
<tr>
<td>Age</td>
<td>-5.664</td>
<td>[-12.37, 1.18]</td>
<td>-0.312*</td>
</tr>
<tr>
<td>Age at diagnosis</td>
<td>-5.187</td>
<td>[-11.86, 1.86]</td>
<td>-0.289*</td>
</tr>
<tr>
<td>Age at menarche</td>
<td>2.996</td>
<td>[-3.77, 9.32]</td>
<td>0.172</td>
</tr>
<tr>
<td><strong>Follicle-stimulating hormone</strong></td>
<td></td>
<td></td>
<td></td>
</tr>
<tr>
<td>Salivary cortisol intercept</td>
<td>1.04</td>
<td>[-6.31, 8.45]</td>
<td>0.233*</td>
</tr>
<tr>
<td>Salivary cortisol slope</td>
<td>0.286</td>
<td>[-5.6, 6.61]</td>
<td>0.031</td>
</tr>
<tr>
<td>Perceived Stress Scale</td>
<td>-1.332</td>
<td>[-2.47, -0.05]</td>
<td>-0.53*</td>
</tr>
<tr>
<td>Hair cortisol</td>
<td>0.381</td>
<td>[-0.93, 1.65]</td>
<td>0.151</td>
</tr>
<tr>
<td>Body mass index</td>
<td>0.294</td>
<td>[-0.99, 1.54]</td>
<td>0.117</td>
</tr>
<tr>
<td>Age</td>
<td>0.153</td>
<td>[-1.22, 1.51]</td>
<td>0.058</td>
</tr>
<tr>
<td>Age at diagnosis</td>
<td>0.113</td>
<td>[-1.32, 1.52]</td>
<td>0.044</td>
</tr>
<tr>
<td>Age at menarche</td>
<td>0.571</td>
<td>[-0.77, 1.7]</td>
<td>0.228*</td>
</tr>
<tr>
<td><strong>Luteinizing hormone</strong></td>
<td></td>
<td></td>
<td></td>
</tr>
<tr>
<td>Salivary cortisol intercept</td>
<td>2.259</td>
<td>[-8.73, 13.95]</td>
<td>0.387*</td>
</tr>
<tr>
<td>Salivary cortisol slope</td>
<td>2.802</td>
<td>[-7.85, 10.18]</td>
<td>0.233*</td>
</tr>
<tr>
<td>Perceived Stress Scale</td>
<td>-0.537</td>
<td>[-2.08, 1.04]</td>
<td>-0.164</td>
</tr>
<tr>
<td>Hair cortisol</td>
<td>0.814</td>
<td>[-0.88, 2.87]</td>
<td>0.247*</td>
</tr>
<tr>
<td>Body mass index</td>
<td>-0.384</td>
<td>[-2.4, 1.65]</td>
<td>-0.117</td>
</tr>
<tr>
<td>Age</td>
<td>-0.43</td>
<td>[-2.48, 1.46]</td>
<td>-0.126</td>
</tr>
<tr>
<td>Age at diagnosis</td>
<td>0.857</td>
<td>[-1.34, 2.87]</td>
<td>0.254*</td>
</tr>
<tr>
<td>Age at menarche</td>
<td>-0.091</td>
<td>[-1.67, 1.55]</td>
<td>-0.028</td>
</tr>
<tr>
<td><strong>Estradiol</strong></td>
<td></td>
<td></td>
<td></td>
</tr>
<tr>
<td>Salivary cortisol intercept</td>
<td>0.14</td>
<td>[-13.51, 14.35]</td>
<td>0.00</td>
</tr>
<tr>
<td>Salivary cortisol slope</td>
<td>1.065</td>
<td>[-13.37, 14.36]</td>
<td>0.001</td>
</tr>
<tr>
<td>Perceived Stress Scale</td>
<td>0.749</td>
<td>[-12.59, 14.75]</td>
<td>0.003</td>
</tr>
<tr>
<td>Hair cortisol</td>
<td>-0.535</td>
<td>[-14.05, 13.34]</td>
<td>-0.002</td>
</tr>
<tr>
<td>Body mass index</td>
<td>0.589</td>
<td>[-13.15, 13.94]</td>
<td>0.002</td>
</tr>
<tr>
<td>Age</td>
<td>-0.1</td>
<td>[-14.03, 13.36]</td>
<td>-0.00</td>
</tr>
<tr>
<td>Age at diagnosis</td>
<td>-0.96</td>
<td>[-14.56, 12.92]</td>
<td>-0.004</td>
</tr>
<tr>
<td>Age at menarche</td>
<td>-1.115</td>
<td>[-14.54, 12.86]</td>
<td>-0.005</td>
</tr>
</tbody>
</table>

*Predictors with the highest standardized regression coefficients
CI—confidence interval
in CCSs, but only in unadjusted models (Barton et al., 2013). The association among perceived stress, HPA activity, and the ovarian reserve has not been examined in CCSs. However, Pal, Bevilaqua, and Santoro (2010) examined the association between AMH levels and chronic psychosocial stress in 89 premenopausal infertile women. The study found that women with chronic stress demonstrated significantly lower AMH levels (p = 0.024) and were three times more likely to be diagnosed with diminished ovarian reserve (p = 0.025). Pal et al. (2010) concluded that chronic psychosocial stress was associated with diminished ovarian reserve. The study proposed HPA dysregulation as a plausible explanation for this association; because a biomarker of chronic stress was not included in the study, the study was unable to provide evidence supporting this theory.

In the current study, the authors found that, with an increase of one standard deviation in hair cortisol concentrations, AMH increased 3.76 pmol/L. Although the standardized regression coefficients of the diurnal cortisol intercept and slope were lower, they were also positively associated with AMH levels. Taken together (positive associations among hair cortisol concentrations, diurnal cortisol intercept and slope, and AMH levels), these findings suggest a positive association between cortisol concentrations and the ovarian reserve, and support the hypothesis that threshold glucocorticoid levels are necessary to sustain ovarian function (Whirledge & Cidlowski, 2013). As stated previously, the authors found lower mean hair cortisol concentrations in the current sample than reported in healthy populations (Stalder & Kirschbaum, 2012).

More than half of the participants in the sample exhibited flattened diurnal cortisol slopes. Lower daily cortisol output and flattened diurnal cortisol slopes have been observed in cancer survivors. These findings are consistent with prior studies linking hypocortisolism to increased risk of experiencing late effects of cancer treatment (Bower et al., 2005; Cuneo et al., 2017; Ho et al., 2013; Schrepf et al., 2015;Sephton et al., 2013).

In this study, the authors found that the model predictors explained 42% of the variation in FSH levels, with the highest standardized regression coefficients observed for salivary cortisol intercept, perceived stress, and age at menarche. The model predictors explained 39% of the variation in LH levels, with the highest standardized regression coefficients observed for salivary cortisol intercept and slope, hair cortisol, and age at diagnosis. The authors also found that perceived stress was inversely associated with FSH and LH levels.

Because the authors were unable to time the collection of reproductive hormone levels according to menstrual cycle phase, a strong association among perceived stress and FSH and LH levels was not anticipated. Several possible explanations exist for the strong inverse association the authors observed irrespective of menstrual cycle phase. It may be that perceived stress has a significant effect on ovarian function regardless of menstrual cycle phase (Schliep et al., 2015). Another explanation may be that stress perception varies across the menstrual cycle, such that the degree to which a woman perceives her life to be stressful is, in part, influenced by fluctuations in gonadotropin levels (Duchesne & Pruessner, 2013).

Although perceived stress was inversely associated with FSH and LH, the authors found that HPA activity (hair cortisol, salivary cortisol intercept, and salivary cortisol slope) was positively associated with FSH and LH levels. The findings support the role of glucocorticoids in regulating the hypothalamic-pituitary-gonadal axis (Whirledge & Cidlowski, 2013). Another explanation is that cortisol levels may fluctuate across the menstrual cycle, as has been suggested by other studies (Kirschbaum, Kudielka, Gaab, Schommer, & Hellhammer, 1999; Nepomnaschy et al., 2011; Stephens, Mahon, McCaul, & Wand, 2016; Wolfram, Bellingrath, & Kudielka, 2011). Although preliminary, the findings from this small sample support an association among perceived stress, HPA activity, and gonadotropin levels in female CCSs. More research is needed to clarify the mechanism behind this association and to determine whether it varies depending on the temporal nature of the stressor and menstrual cycle phase.

Limitations
This exploratory study has several limitations. Because of the small sample, the authors were unable to adjust for hormonal contraceptive use in the overall model. Hormonal contraceptive use is associated with suppression of FSH and LH levels, as well as increased production of corticosteroid-binding globulin and subsequent decreases in unbound cortisol levels.

KNOWLEDGE TRANSLATION
- Perceived stress should be assessed in female childhood cancer survivors (CCSs) who report reproductive issues.
- Hair cortisol may provide a convenient, noninvasive means of assessing hypothalamic–pituitary–adrenal activity in CCSs.
- Providers should consider factors other than diagnosis and treatment-related variables when counseling CCSs regarding their future fertility.
Further research with larger sample sizes is needed to control for the effects of contraceptive use on ovarian function and HPA activity in female CCSs. In addition, because of the wide variety of pediatric cancer diagnoses in this small sample, the authors could not adjust for cancer diagnosis- and treatment-related factors in the model. Several studies have found a significant effect of diagnosis- and treatment-related factors on post-treatment ovarian function, including the cancer pathology itself (van Dorp et al., 2014), the therapeutic regimen (Charpentier et al., 2014), and the length of exposure (El-Shalakany, Ali, Abdelmaksoud, Abd El-Ghany, & Hasan, 2013; Reinmuth et al., 2013). Another limitation is that, because of the great distance that many of the participants needed to travel, the authors were unable to time study visits to coincide with a specific menstrual cycle phase. For this reason, the authors were unable to adjust gonadotropin and reproductive hormone levels to account for menstrual cycle phase. Lastly, the study design was cross-sectional and relied on a convenience sample. The study’s findings require validation in larger longitudinal studies.

Implications for Nursing
Reproductive dysfunction is a major concern for cancer survivors and is highly correlated with quality of life in this vulnerable population (Cherven et al., 2015; Knopman et al., 2010; Kondapalli et al., 2014; Letourneau et al., 2013). However, studies have demonstrated that cancer survivors are about half as likely to be treated for infertility as their siblings (Barton et al., 2013). Although the reasons for this discrepancy are not fully understood, healthcare providers may perceive fertility treatments to be less successful in this population. The guidelines from the American Society for Clinical Oncology recommend that options for fertility preservation be discussed with patients with cancer at the earliest opportunity (Knight et al., 2015; Letourneau et al., 2013). Oncology nurses play an important role in advocating for cancer survivors, helping to initiate conversations regarding future fertility as early as possible, and discussing factors that may mitigate the risk for post-treatment reproductive dysfunction. The findings from this study suggest that there may be modifiable factors, such as chronic stress, that contribute to risk for infertility post-treatment. Oncology nurses can minimize the potential burden of late effects by educating patients regarding their risk and what actions can be taken prior to or after treatment to reduce this risk (McClellan et al., 2013).

Conclusion
In this exploratory study, the authors sought to examine the association between chronic stress and ovarian function in survivors of childhood cancer. Although there is prior evidence suggesting stress affects gonadotropin hormone levels, the effects of chronic stress on the ovarian reserve and its biomarker AMH are poorly understood. The findings from this study provide preliminary evidence to suggest that perceived stress is negatively associated with ovarian function and that biomarkers of HPA activity are positively associated with ovarian function in female CCSs. Few studies have examined predictors of the ovarian reserve in CCSs apart from diagnosis- and treatment-related factors. The findings from this study provide the foundation for further research examining the risk for post-treatment reproductive dysfunction in female CCSs.

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