A Gene Expression Screen Identifies **EARLI1** as a Novel Vernalization-Responsive Gene in *Arabidopsis Thaliana*

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Vernalization promotes early flowering in late ecotypes of \textit{Arabidopsis thaliana}. The mechanisms of vernalization are poorly understood. A subtractive hybridization approach was used to isolate vernalization-responsive genes from a late-flowering ecotype of \textit{Arabidopsis thaliana} based on the premise that transcript levels of such genes would increase with cold treatment and remain high even after removal of the vernalization stimulus. \textit{EARLI1} is the first Arabidopsis gene shown to be stably activated by vernalization. The abundance of its RNA is progressively elevated by vernalization and remains high for at least 20 days at room temperature. The basal level of \textit{EARLI1} RNA is higher in early-flowering ecotypes, but is increased also after vernalization. Vernalization and subsequent growth in long-day photoperiods have an additive or synergistic effect on \textit{EARLI1} activation. \textit{EARLI1} RNA levels are also transiently induced by brief exposures.
to cold, but not to abscisic acid. EARLI1 is thus a novel vernalization-responsive gene in Arabidopsis thaliana that can be used to investigate vernalization-specific transcriptional regulation.

Introduction

The transition from vegetative to reproductive development is a critical stage in the life cycle of plants. The timing of this transition is carefully regulated to synchronize flowering with environmental conditions that ensure maximum survival of the next generation. Vernalization (prolonged exposure to low temperatures) during winter months followed by long-day photoperiods in the spring promote early flowering in many naturally occurring ecotypes of Arabidopsis thaliana (Martínez-Zapater and Somerville, 1990; Bagnall, 1993; Lee and Amasino, 1995). Seeds then germinate during the summer or fall, but plants remain vegetative until vernalization occurs again over the winter and induces synchronized flowering in the spring. Such Arabidopsis ecotypes are late-flowering without vernalization, because they contain dominant alleles of FRIGIDA (FRI) and FLOWERING LOCUS C (FLC), which synergistically confer the strong requirement for vernalization to induce early flowering (Lee et al., 1993, 1994; Clark and Dean, 1994; Koornneef et al., 1994; Sanda et al., 1997).

The transition to flowering in Arabidopsis is regulated by four interacting genetic pathways (Koornneef et al., 1998; Levy and Dean, 1998; Piñeiro and Coupland, 1998; Simpson et al., 1999). A floral repressor pathway is proposed to maintain the vegetative phase until the flowering promotion pathways initiate reproductive development (Martínez-Zapater et al., 1994). These include the long-day, autonomous, and vernalization promotion pathways of flowering (Koornneef et al., 1998; Levy and Dean, 1998; Simpson et al., 1999). In the absence of dominant alleles of FRI, plants do not require vernalization and flower early due to activities of the autonomous and long-day promotion pathways. However, both dominant FRI and recessive mutations in genes of the autonomous promotion pathway appear to effect late flowering by increasing the level of FLC RNA, suggesting that FLC, a MADS box protein, may be a major repressor of early flowering in Arabidopsis thaliana (Sanda et al., 1996; Michaels and Amasino, 1999; Sheldon et al., 1999, 2000). Conversely, vernalization decreases the level of FLC RNA in late-flowering plants, which correlates with early flowering. We thus define FLC as the first
member of a group of *bona fide* vernalization-responsive genes in *Arabidopsis thaliana*, whose expression responds negatively to vernalization. That is, *FLC* is silenced by vernalization and remains off even after the vernalization stimulus is removed.

This study describes the first example of another group of vernalization-responsive genes in *Arabidopsis thaliana*, those whose expression responds positively to vernalization. Because a drug- or transgenemediated reduction of cytosine methylation partially substituted for vernalization in late-flowering plants, it was proposed that vernalization may activate genes by de-methylation of specific DNA sequences (Burn *et al*., 1993; Dennis *et al*., 1996, 1997; Finnegan *et al*., 1998). Using a traditional subtractive hybridization approach, we were able to identity *EARLI1* as a *bona fide* vernalization-responsive gene in *Arabidopsis thaliana*, because the gene is activated by vernalization and its RNA abundance remains high even after the vernalization stimulus is removed. We show here that the level of *EARLI1* RNA is progressively elevated with increased length of vernalization and that it remains high for at least 20 days after vernalization. *EARLI1* transcript levels are further increased when plants are grown in long-day photoperiods after vernalization. *EARLI1* is also transiently induced by a short period of cold, but not by abscisic acid. Hence we define *EARLI1* as a novel vernalization-responsive gene.

**Materials and methods**

**Plant material and treatment**

Early-flowering *Arabidopsis* ecotypes Columbia (Col0) and Nossen (No-0) were obtained from ABRC (Ohio State University, OH) and Landsberg erecta (Ler) was kindly provided by T.-P. Sun (Duke University, NC). Late-flowering ecotypes containing dominant alleles of *FRIGIDA* (*FRI*) and *FLOWERING LOCUS C* (*FLC*) in the Col-0 background were kindly provided by E. Himelblau and R. Amasino (University of Wisconsin-Madison, WI; Lee *et al*., 1993). About 500 surface-sterilized seeds were grown *in vitro* in a sterile petri dish (9 cm diameter) on agar-solidified half-strength MS medium without sucrose (Murashige and Skoog, 1962). Unless otherwise stated, all plates
containing unvernalized control seeds were placed at 4°C for 2 to 3 days to break seed dormancy, then grown for 7 days with a 8 h short-day photoperiod under cool fluorescent light and ca. 20°C day/night temperature. Unless otherwise stated, seeds were vernalized on agar plates at 4°C for 14 to 35 days in 8 h short-day photoperiods under cool fluorescent light, then grown for 7 days with a short-day photoperiod and 20°C day/night temperature. For transient exposure to cold, seeds were grown after breakage of dormancy on agar medium for 7 days at 20°C in an 8 h short-day photoperiod, then transferred overnight to 4°C and then grown for 0 to 8 h at 20°C. For transient exposure to abscisic acid (ABA), 10 ml of a 100 μM solution of ABA (Sigma) in 1% DMSO was added to plates with 7-day old seedlings grown in short-day photoperiods. Seedlings were incubated for 5 min, then the ABA was washed away. Plates were then left in the light for 2 to 8 h at 20°C. Control plates were treated the same way with 1% DMSO lacking ABA.

**RNA isolation**

Unless otherwise stated, whole-seedling tissue was harvested by quick freezing in liquid N2 and stored at −70°C. RNA from vernalized and unvernalized tissues was isolated at the same time. The frozen tissue was ground to a fine powder on dry ice in a mortar and pestle with added liquid N2. Total RNA was isolated by a modified miniprep procedure as described previously (Yeh et al., 1990). Briefly, 100–500 mg of crushed tissue was resuspended in 1 ml of extraction buffer (7.5 M guanidine hydrochloride (Sigma), 25 mM sodium citrate (Sigma), 0.5% w/v sodium lauryl sarcosine (Sigma), 0.1 M 2-mercaptoethanol) and spun for 10 min at maximum speed in an Eppendorf centrifuge. The supernatant was extracted three times with an equal volume of phenol/chloroform/isoamyl alcohol (24:24:1), followed by one extraction with chloroform/isoamyl alcohol (24:1), and total RNA was precipitated at −20°C in an equal volume of isopropanol and 0.1 volume of 3 M sodium acetate (Sigma). The pellet was washed in 75% v/v cold ethanol, vacuum-dried, and resuspended in 30–50 μl DEPC-treated water. Total RNA was quantified spectrophotometrically and stored in aliquots (20 μg) at −70°C.
Subtractive hybridization and colony screening

Three rounds of reciprocal subtractive hybridization were performed as described previously (Wang and Brown, 1991; Cook et al., 1995). Briefly, poly(A)+ mRNA from vernalized and unvernalized plants was selected by passage of total RNA through an oligo(dT) cellulose column from the Oligotex mRNA Mini Kit (Qiagen) and cDNA was synthesized with a cDNA synthesis kit as recommended by the supplier (Amersham). Double-stranded cDNA was dissolved in 20 μl water and 8 μl aliquots were digested with either AluI or AluI plus RsaI (American Allied Biochemical) and ligated to a duplex oligonucleotide containing one blunt end and a 4 base protruding 3' end. The duplex oligonucleotide was made by mixing the kinase-digested 25-mer oligonucleotide 5'TAGTCCGAATTCCAAGCAAGGCACA-3' with the 21-mer oligonucleotide 5'-CTCTTGCTTGAATTCCGGACTA-3' (EcoRI sites in bold). The ligated cDNA was purified from unligated linker by electrophoresis through a 1.4% low-melting agarose gel. The region containing fragments of 150 bp to 1.5 kb was cut from the gel and stored at 4°C. The gel slices were dissolved at 65°C and 1 μl was used for 100 μl standard PCR reactions in 1× buffer (Promega) containing 1.25 mM MgCl2, 1 μg of the 21-mer primer, 200 μM dNTPs, and 2.5 units of Taq polymerase (Promega). PCR-amplified driver cDNA (55 μg) was cut with EcoRI and biotinylated twice with 50 μg Photobiotin (Vector Labs) in an ice bath under a 275 W sunlamp (Vector Labs). For each round of subtractive hybridization, 1.25 μg of tracer cDNA was denatured together with 25 μg of biotinylated driver cDNA at 100°C and hybridized for 20 h (long hybridization) in 1.5 M NaCl, 50 mM Hepes pH 7.3, 10 mM EDTA, 0.2% SDS in a 68°C water bath. Biotinylated driver and tracer cDNA that was hybridized to driver cDNA was removed by repeated addition of 10–15 μg streptavidin and phenol/chloroform extractions, then incubated for 2 h (short hybridization) with 12.5 μg of biotinylated driver cDNA, followed by streptavidin/phenol/chloroform extractions. After each round, subtracted cDNA was PCR-amplified as described and used in the next round of subtractive hybridization. After three rounds, cDNA fragments were cloned into EcoRI-digested pBlueScript II plasmid (Stratagene).

A reverse northern hybridization method was used to specifically screen for cDNA fragments of genes that were induced or up-regulated
by vernalization. Individually cloned, subtracted cDNA fragments from either vernalized or unvernalized plants were PCR-amplified with pBlueScript plasmid vector-specific primers and slot-blotted (1 μl) in duplicate onto nitrocellulose membranes using the Bio-Dot SF Microfiltration Apparatus (BioRad). Individual membranes containing subtracted cDNA fragments from vernalized and unvernalized plants were hybridized with first-strand cDNA made from RNA of either vernalized or unvernalized plants. This reverse northern first-strand cDNA probe was prepared in the presence of 10 μCi of α32P-dATP using a cDNA synthesis kit (Amersham) as recommended by the supplier. Colonies that only hybridized to the cDNA probe made from RNA of vernalized plants were selected and used in RNA gel blot analyses.

**RNA gel blot analysis and probes**

About 20 μg of total RNA was separated by electrophoresis in 1.2% formaldehyde gels containing ethidium bromide (Sambrook et al., 1989). RNA was transferred onto nitrocellulose membranes and cross-linked with a Stratalinker (Stratagene). Prehybridization and hybridization was performed at 65°C in 5× Denhardt’s solution, 6× SSC, 0.5% SDS and 0.1 mg/ml denatured salmon sperm DNA. All probes were 32P-labeled by the random primer method using the MegaPrime labeling kit as recommended by the supplier (Amersham), purified by G-50 spin columns, heat-denatured and hybridized at a concentration of 1 × 10⁶ cpm/ml onto nitrocellulose membranes in a Hybaid oven (Labnet) at 65°C for at least 16 h. Membranes were washed twice in 2× SSC/1% SDS for 5 min at room temperature, then in 0.2× SSC/0.1% SDS for at least 30 min at 65°C. Membranes were briefly rinsed in 0.2× SSC before autoradiography.

Unless otherwise stated, EARLI1 probes were made from the 208 bp cDNA fragment isolated by subtractive hybridization (see Figure 2). A full-length EARLI1 probe was made from cDNA generated by RT-PCR with Superscript II (Gibco-BRL) and Taq polymerase (Promega) using primers 5‘-ACTGAATGGATCCTTAAAACAAAC-3‘ and 5‘TAGCTGAATTCCTTCAAGCACATT-3‘. FLC probes were made from cDNA lacking the conserved MADS box region (kindly provided by S. Michaels and R. Amasino, University of Wisconsin-Madison). **COR15α**
(Lin and Thomashow, 1992) and ACT2/ACT8 (An et al., 1996) probes were made from 3'-UTR regions of cDNA isolated by RT-PCR with COR15α primers 5'-AGATTTGCAGCAGGATAAA-3' and 5'-TGTGACGTTGACTGGATA-3' and ACT2/ACT8 primers 5'-GCTTCGTATTCCTGA-3' and 5'-TAGCTGAATTCTTCAAGCATT-3'.

Results

**EARLI1 is a vernalization-responsive gene that is activated in all Arabidopsis tissues**

A sensitive PCR-based gene expression screen was used to identify vernalization-responsive genes in late-flowering *Arabidopsis thaliana* containing dominant alleles of *FRI* and *FLC* (see Materials and methods). By definition, the RNA level of vernalization-responsive genes is either stably increased or decreased by vernalization, which is distinct from the transient activation of cold acclimation or stress-responsive genes (Thomashow, 1994). To identify *bona fide* vernalization-responsive genes in *Arabidopsis*, seeds imbibed on agar medium were vernalized for 35 days at 4°C, then grown at room temperature for seven days prior to RNA isolation. The seedlings had two cotyledons and two leaves at this stage. Unvernalized, 2-day cold-treated, control plants were grown in parallel and were phenotypically identical to vernalized plants after 7 days at room temperature. All plants were grown in short-day photoperiods to reduce differences in long-day photoperiod-specific gene expression between vernalized and unvernalized plants.

The MADS box gene *FLC* is the first known vernalization-responsive gene whose RNA level is decreased by vernalization in *Arabidopsis thaliana* (Michaels and Amasino, 1999; Sheldon et al., 1999). Therefore, *FLC* served as positive control in repeated rounds of reciprocal subtractive hybridization between cDNA from vernalized and unvernalized *Arabidopsis* plants. *FLC* was indeed selectively and progressively enriched in cDNA from unvernalized plants during three rounds of subtractive hybridization, but was absent in cDNA from 35-day vernalized plants (data not shown). Conversely, the amount of the constitutively expressed actin gene *ACT2/ACT8* was progressively reduced in cDNA from both vernalized and unvernalized plants during
three rounds of subtractive hybridization (data not shown). Hence it was concluded that both the cDNA material and the experimental approach were suitable for the identification of novel, vernalization-responsive genes.

After three rounds of subtraction against cDNA from unvernalized control plants, PCR-amplified cDNA fragments from vernalized plants were ligated to plasmid vectors and cloned. About 100 individual clones were hybridized in duplicate with radiolabeled probes made from the two original, unsubtracted cDNA populations. Two clones were identified that only hybridized to cDNA from vernalized plants. Both clones had ca. 200 bp identical inserts and were used in RNA gel blot analysis with total RNA from vernalized and unvernalized plants grown for seven days at room temperature in short-day photoperiods. As shown in Figure 1, the FLC control hybridized strongly to RNA from only unvernalized, but not vernalized, plants. When the same blot was stripped and re-probed with the 200 bp cDNA fragment, hybridization signal was only seen in RNA from vernalized plants. Thus, the gene expression screen identified a novel vernalization-responsive gene that is strongly activated by 35 days of vernalization, and remains on after 7 days of growth in short-day photoperiods.

The ca. 200 bp cDNA fragment was sequenced and analyzed. As shown in Figure 2A, database searches determined that the 208 bp cDNA fragment was identical to EARLI1 (accession number L43080), originally identified as a gene transiently activated in response to toxic levels of aluminum (Richards and Gardner, 1995). EARLI1 belongs to a group of small (ca. 18.7 kDa), hydrophobic, and proline-rich proteins that are developmentally and environmentally regulated in a wide variety of plant species (Richards and Gardner, 1995). As shown in Figure 2A, EARLI1 contains a putative signal sequence at the N-terminus, followed by a proline-rich hydrophilic domain, and a hydrophobic C-terminus with a putative transmembrane domain. EARLI1 maps to Arabidopsis chromosome 4, between RFLP markers g4108 and mi456, and is located on BAC T1P17 (accession number AL049730). Database analysis of BAC T1P17 showed that EARLI1 is flanked by five related EARLI1-like genes, which are tandemly repeated over a region of 20 kb. The genes contain intron-less coding
regions and they are separated by 2 to 3 kb of 5'-and 3'-flanking sequences. Another related protein was found on BAC T3P18 that maps to chromosome 1. A protein alignment (not shown) indicated that the highest degree of similarity between the related proteins exists at the C-terminus. As shown in the nucleotide alignment in Figure 2B, the 208 bp \textit{EARLI1} cDNA fragment maps to a region of low similarity between the most related genes. When the 208 bp fragment was used in a genomic Southern hybridization experiment, only bands corresponding to \textit{EARLI1}-specific restriction fragments were detected (not shown). We thus concluded that the hybridization signal seen in RNA gel blot analyses was specific for \textit{EARLI1}.

The expression of \textit{EARLI1} shown in Figure 1 was analyzed in RNA isolated from whole seedlings. To determine whether \textit{EARLI1} activation was tissue-specific in vernalized plants, total RNA was isolated from individual tissues and subjected to gel blot analysis. \textit{EARLI1} transcripts were detected at approximately equal levels in the shoot apex, cotyledons, the hypocotyl, and in roots (data not shown). This indicated that \textit{EARLI1} is activated in all tissues tested and suggested that the gene is under developmental, but not spatial, control.

\textit{Extended periods of vernalization quantitatively activate stable EARLI1 expression}

The vernalization response is quantitative. That is, the longer plants are vernalized, the earlier they flower (Lee and Amasino, 1995). One prediction from this observation is that \textit{bona fide} vernalization-responsive genes are also quantitatively activated or silenced during extended periods of vernalization. To test this prediction, the level of \textit{EARLI1} and \textit{FLC} transcripts was measured in plants exposed to varying periods of vernalization treatment. As before, all plants were grown for seven days at room temperature and in short-day photoperiods prior to RNA isolation. As shown in Figure 3A, \textit{EARLI1} RNA levels rapidly and stably increased with every week of vernalization, reaching the highest level in 35-day vernalized plants. In contrast, \textit{FLC} RNA levels were highest in unvernalized plants, but rapidly and stably decreased with every additional week of vernalization to almost undetectable levels (Figure 3A). Thus, both \textit{EARLI1} and \textit{FLC} satisfy the conditions for \textit{bona
fide vernalization-responsive genes, whose transcripts are quantitatively induced and repressed, respectively.

To determine how long EARLI1 expression levels remain high after vernalization, 35-day vernalized plants were grown for 7, 10, 15, and 20 days at room temperature. No decline in level of EARLI1 transcripts was seen over a 10-day growth period and mRNA levels were still high after 20 days (Figure 3B). Thus, the activated state of EARLI1 gene expression was maintained for at least 20 days of growth at room temperature.

**Basal EARLI1 expression is higher in early-flowering ecotypes**

We observed an inverse correlation between RNA levels of EARLI1 and FLC as a function of vernalization (Figure 3A). This suggested that FLC may repress EARLI1. If so, it might be expected that the basal level of EARLI1 RNA is higher in early-flowering plants, which contain a very low basal level of FLC RNA. EARLI1 mRNA abundance was thus analyzed in three early-flowering ecotypes of *Arabidopsis*, Col-0, No-0, and Ler. To measure basal or elevated transcript levels, seeds were kept for two days in the cold or vernalized for 28 days, then grown for seven days at room temperature in short-day photoperiods. The result of RNA gel blot analyses is shown in Figure 4. As in late-flowering plants, EARLI1 is activated and remains on after 28 days of vernalization in early-flowering plants. However, compared to unvernalized late-flowering plants, the basal level of EARLI1 RNA is clearly higher in early-flowering ecotypes, particularly in Ler, which contains a weak allele of FLC (Lee et al., 1994; Koornneef et al., 1994). Thus, FLC may indeed directly or indirectly repress EARLI1 expression.

**Vernalization coupled with long-day photoperiods have an additive or synergistic effect on EARLI1 expression**

*EARLI1* is efficiently activated when plants are both vernalized and grown in short-day photoperiods (Figure 1). To determine whether *EARLI1* expression is responsive to differences in photoperiod, four
pools of seeds were vernalized for either 14 or 28 days in either long- or short-day photoperiods. Subsequently, each pool of vernalized seeds was transferred to room temperature, further subdivided, and grown separately in either long-or short-day photoperiods. Hence, some plants were vernalized in short days, then grown in either long- or short-day photoperiods for one week before RNA isolation. Conversely, other plants were vernalized in long days, then grown in long- or short-day photoperiods before RNA isolation. As shown in Figure 5, the level of EARLI1 RNA was always higher in plants that were grown in long-day photoperiods after vernalization, regardless of the photoperiod during vernalization. Although EARLI1 was activated after vernalization and growth in short days, vernalization followed by growth in long-day photoperiods had an additive or synergistic effect on gene activation. Thus, vernalization may make EARLI1 competent to respond to long-day photoperiods.

**EARLI1 is not a general stress-responsive gene**

*EARLI1* was previously identified as a gene whose RNA was transiently induced by exposure to toxic levels of aluminum (Richards and Gardner, 1995). This suggests that *EARLI1* may be transiently activated by abiotic stress, as shown for other proline-rich proteins (Kurkela and Franck, 1990). To test this prediction, unvernalized 7-day old plants were exposed only overnight to cold, then grown for 0 to 8 h at room temperature prior to RNA isolation. In another experiment, unvernalized 7-day old plants were exposed for 5 min to 100 μM of the stress hormone abscisic acid (ABA) to determine whether ABA was able to transiently activate *EARLI1*. The level of *EARLI1* expression was measured at each time point by RNA gel blot analysis and compared to the level of *COR15α* expression. *COR15α* served as positive control, because its RNA level is transiently elevated by both cold and ABA (Hajela *et al.*, 1990; Thomashow, 1994). As shown in Figure 6, *EARLI1* was transiently activated by cold, but not by exposure to ABA. By contrast, *COR15α* was activated by both cold and ABA. Both *EARLI1* and *COR15α* transcripts returned to basal levels with similar kinetics after removal of transient cold, suggesting that the stable activation of *EARLI1* by vernalization is promoted by a different mechanism. Thus, the regulation of *EARLI1* is distinct from that of *COR15α*, a cold acclimation-responsive gene, in three ways. First, *EARLI1* is stably
activated by vernalization and COR15α is not (Thomashow, 1994). Second, EARLI1 is transiently activated only by cold and not by ABA, whereas both treatments induce COR15α. And third, EARLI1 does not have a COR15α-like low-temperature-responsive element (LTRE) in its promoter sequence (data not shown), suggesting that another control element may respond to transient cold. We hence conclude that EARLI1 is a novel vernalization-and cold-responsive gene with a unique expression pattern distinct from general stress-responsive genes.

**Discussion**

*Identification of EARLI1 as a vernalization responsive gene in Arabidopsis thaliana*

The paucity of vernalization-responsive genes has been a major obstacle to analyzing vernalization-specific regulation of gene expression in Arabidopsis thaliana. Previous to this study, the only cloned Arabidopsis gene that fits the definition of bona fide vernalization-responsive gene was FLOWERING LOCUS C (FLC; Michaels and Amasino, 1999; Sheldon et al., 1999). The level of FLC RNA is high in unvernalized, FRIGIDA (FRI)-containing late-flowering plants, but is decreased after vernalization and remains low even after removal of the vernalization stimulus (Michaels and Amasino, 1999; Sheldon et al., 1999; this study). The observed expression profile fits FLC's proposed function as a MADS box gene encoding a transcription factor that represses flowering (Simpson et al., 1999). We report here on the identification of EARLI1, the first vernalization-responsive gene from Arabidopsis that is activated by vernalization and remains on after removal of the cold stimulus. The abundance of EARLI1 mRNA is very low in unvernalized, late-flowering Arabidopsis plants, but is robustly increased and remains high after vernalization in all vegetative tissues tested. This up-regulation is quantitative, thus the longer plants are vernalized, the higher the abundance of EARLI1 mRNA. Conversely, we found that the level of FLC mRNA is quantitatively decreased to very low levels over a 35-day period of vernalization (Figure 3). The inverse RNA profiles of EARLI1 and FLC may also correlate with the quantitative effect of vernalization on flowering time (Lee and Amasino, 1995). That is, the longer late
ecotypes of Arabidopsis are vernalized, the earlier they flower, which correlates with higher and lower levels of EARLI1 and FLC RNA, respectively (Schläppi, unpublished; Sheldon et al., 2000). It is thus possible that FLC directly or indirectly represses EARLI1 expression. Consistent with this idea, we found that, compared to late-flowering plants, EARLI1 is more highly expressed in unvernalized, early-flowering ecotypes of Arabidopsis such as Col-0 or Ler (Figure 4). This would suggest that EARLI1 is under negative control by FLC. However, we consider it equally or more likely that EARLI1 is mainly under positive control by other factors. This is because vernalization activates the gene even in the Ler ecotype, which has undetectable levels of FLC RNA (Figure 4; Sheldon et al., 1999). Moreover, typical consensus sequence binding sites for MADS-box proteins (Riechmann and Meyerowitz, 1997) are not found in the 5′-upstream region of EARLI1 (not shown), which may suggest more likely an indirect effect of FLC.

In this initial characterization we present evidence that EARLI1 is a bona fide vernalization-responsive gene in Arabidopsis that is specifically activated by vernalization. The only vernalization-related (ver) genes other than EARLI1 that have been reported to date were found in winter wheat (Chong et al., 1994, 1998). Like EARLI1, vernalization-related winter wheat genes were induced after 26 to 30 days of vernalization, but it was not reported as to whether transcript levels remain high after removal of the vernalization stimulus.

**EARLI1 is regulated by both vernalization and long-day photoperiods**

The level of EARLI1 transcript is higher in vernalized plants grown in long days than in vernalized plants grown in short days (Figure 5). In contrast, the basal level of EARLI1 transcript is similar in unvernalized, late-flowering ecotypes grown in short- or long-day photoperiods (data not shown). Taken together, this suggests that vernalization makes EARLI1 competent to respond to long-day photoperiods. It is thus possible that the EARLI1 promoter is modular, containing both novel vernalization and long-day-responsive elements that act additively or synergistically to increase RNA levels. Since EARLI1 is intron-less (Figure 2), it seems likely that the gene is mainly
regulated by its 5' control region. The 5'-upstream region of EARLI1 contains several myb core elements and different myb consensus sites (not shown), which may be candidate sequences for photoperiodic regulation (Wang et al., 1997; Schaffer et al., 1999). In addition, several GT-1 consensus binding sites involved in light regulation (Terzaghi and Cashmore, 1995) are also found in the same 5' upstream regions (not shown). Vernalization response elements are unknown and most likely represented by novel DNA sequences. Analysis of changes in the DNA methylation state of EARLI1 promoter elements before and after vernalization may help to identify vernalization-responsive elements. Alternatively, different response elements could be identified by traditional promoter deletion analysis.

DNA sequence motifs CCGAC for the low-temperature response elements (LTRE; Baker et al., 1994) are not present in the 5' upstream region of EARLI1 (not shown). However, EARLI1 RNA is transiently induced by overnight exposure to cold, but not by the stress hormone abscisic acid (ABA), which is distinct from LTRE-mediated regulation (Figure 6). It is thus possible that the EARLI1 promoter contains both vernalization and novel low-temperature-responsive elements, the latter only responding to transient cold and not to ABA-mediated, general stresses. However, the possibility that EARLI1 is also responsive to other types of abiotic stresses such as high salt or wounding needs to be addressed in further experiments.

Possible function of EARLI1

In this study we provide evidence that EARLI1 is activated by vernalization, but we do not know whether the gene is required for vernalization-promoted early flowering. The EARLI1 protein belongs to a class of small proline-rich proteins found in a variety of plant species (Richards and Gardner, 1995) with similarity to extensin-like or putative cell wall plasma membrane-disconnecting CLCT proteins in Arabidopsis (e.g. accession number AAC22151 and AF098630, respectively). The protein may thus be plasma membrane-or cell wall-anchored and could be involved in signal transduction pathways. Alternatively, it could affect the developmental potential of certain cells by modifying plasma membrane or cell wall properties. In addition to
testing whether *EARLI1* is involved in the regulation of flowering time, it will be interesting to determine whether *EARLI1* is regulated by known genes of the vernalization promotion pathway (Chandler *et al.*, 1996).

In summary, we have identified a vernalization-responsive gene in *Arabidopsis thaliana* that responds positively to vernalization and long-day photoperiods, to transient cold, but not to ABA. It should now be possible to identify cis-acting control elements that respond positively to the different environmental stimuli. This may ultimately allow us to identify vernalization-specific trans-acting factors and to advance the understanding of the relatively obscure process of vernalization-promoted flowering.

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Appendix

Figure 1
Vernalization-responsive expression of a gene identified by subtractive hybridization. Upper panel: steady-state mRNA abundance of FLC (Michaels and Amasino, 1999) is stably decreased in the late-flowering, FRIGIDA-containing Columbia ecotype of Arabidopsis thaliana. RNA was prepared from unvernalized (-) and 35-day vernalized (+) seedlings grown in vitro for 7 days at room temperature in short-day photoperiods. The 30 end of FLC cDNA lacking the conserved MADS box sequence was used as a probe to detect FLC mRNA. Middle panel: steady-state mRNA abundance of the identified gene is stably elevated in the FRIGIDA-containing Columbia ecotype after 35 days of vernalization. The same RNA blot shown in the upper panel was reprobed with a 208 bp cDNA fragment isolated by subtractive hybridization to detect a ca. 850 nt vernalization-responsive transcript. Lower panel: the RNA blot was reprobed a second time with the actin gene ACT2/ACT8 (An et al., 1996) to determine the relative amount of RNA loaded in each lane.
Figure 2

Nucleotide sequence, deduced amino acid sequence, and partial nucleotide sequence alignment of the vernalization-responsive gene identified by subtractive hybridization.

A. The gene corresponds to EARLI1 (Richards and Gardner, 1995). EARLI1 is intronless and the genomic sequence is shown. The 208 bp cDNA sequence isolated by subtractive hybridization is underlined. A putative N-terminal signal sequence of the EARLI1 protein is shown in *italics*. The proline-rich domain of the protein is shown in **bold**. B. Partial nucleotide sequence alignment of EARLI1 with four of its most related family members. T1P17.70 corresponds to EARLI1. The cDNA sequence isolated by subtractive hybridization is underlined.

A.

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atg gct tca aag aac tca gcc tct att gct ctt ttc gcc ctt aac atc ata ttc ttc
MAS KNS AS S S T S A L FF F A L N I T F F
acc tta acc gct gcc acc gat tgg ggt tgc acc cca ggt cct aag cac aag cct gtc cca
tL T A A T A D C G C N P S F K K P V P
ag tct atc acc ccg aag cct gcc ACC ggt aac ccc agc gtt cca gat cct gca agt ctg cca gtt ctc gta
SP K K P V P V P K P V P K P V P
/ccg ggt cct tgg gtc cca gat cct aac gct agg cgg gtc acc cct ccg aga acc cct gcc
P S P S V I P S P P V P P T P R T P G
ttc tct cga aac tgt cct atc gat gct ctc aga ctc gtt gta tgt ggc aac gtt ttc aoc
SS G N C P I D A L R L G V C A N V L S
agt cta ctc aac att cca tgt ggt cag cca tca gct cta acc cca tgt tgc ctc atc cca
S L L N I Q L G Q S A Q P C C S L I Q
ggt tgt gtt ggc ctc gcc gct gct acc att tgt ttc gtc act ggc ctt agg gct aac gtt ctt
G L V D L D A A I C L C T A L R A N V L
gtt atc aac ctt aac gcc ata tct ctc agg gtt ctt ctc aac gtt tgt aac aag aag
G I N L N V P I S L S V L L N V C N R K
gtt cgc tct ggc ttc cca tgt gct
V P S G F Q C A
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B.
Figure 3
Quantitative effect of vernalization on EARLI1 and FLC steady-state mRNA levels and postvernalization-sustained expression of EARLI1. A. Upper panel: EARLI1 is progressively activated with increased time of vernalization. RNA was prepared from FRIGIDA-containing late-flowering Columbia seedlings vernalized in vitro for the indicated time periods, then grown for 7 days in short-day photoperiods. The RNA gel blot was simultaneously probed with the 208 bp subtracted cDNA fragment (Figure 2) to detect EARLI1 mRNA and with the actin gene ACT2/ACT8 (An et al., 1996) to determine the relative amount of RNA loaded in each lane. Lower panel: FLC is progressively silenced with increased time of vernalization. RNA was from the same isolation as in the blot shown in the upper panel. The RNA gel blot was simultaneously probed with FLC cDNA (Michaels and Amasino, 1999) lacking the conserved MADS box to detect FLC mRNA and with ACT2/ACT8 to determine the relative amount of RNA loaded in each lane. B. RNA gel blot showing sustained levels of EARLI1 mRNA. Upper panel: RNA was prepared from 35-day vernalized FRI-containing late-lowering Arabidopsis seedlings grown after vernalization for the indicated periods of days at room temperature and in short-day photoperiods. Lower panel: ethidium bromide-stained rRNA (28S) as loading control.
Figure 4
The basal level of EARLI1 steady-state mRNA is higher in unvernalized, early-flowering ecotypes of Arabidopsis thaliana. RNA was prepared from unvernalized (−) or 28-day vernalized (+) seedlings grown in vitro for 7 days at room temperature in short-day photoperiods. The 208 bp subtracted cDNA fragment (Figure 2) was used as a probe to detect EARLI1 mRNA. The RNA gel blots were reprobed with the actin gene ACT2/ACT8 (An et al., 1996) to determine the relative amount of RNA in each lane.

Flowering time of ecotypes in long-day photoperiods is indicated as the number of rosette leaves (RLN) made at the time of bolting. No-0, early-flowering ecotype Nossen; Col-0; early-flowering ecotype Columbia; Col(FRI), late-flowering ecotype containing the Sf-2 allele of FRIGIDA in the Col-0 background (Lee et al. 1993); Ler, early-flowering ecotype Landsberg erecta; n.d., not determined. All early-flowering ecotypes contain recessive frigida (fri) alleles.

Figure 5
Growth in long-day photoperiods after vernalization increases the level of EARLI1 steady-state mRNA. Late-flowering FRIGIDA-containing Columbia seedlings were vernalized in vitro for 14 or 28 days in either short days or long days, then subdivided and grown at room temperature as indicated in either short-day (S) or long-day (L) photoperiods. The 208 bp subtracted cDNA fragment (Figure 2) was used as a probe to detect EARLI1 mRNA. The RNA gel blots were reprobed with the actin gene ACT2/ACT8 (An et al., 1996) to determine the relative amount of RNA in each lane.
Figure 6  
Effect of transient cold and exogenous application of abscisic acid (ABA) on the levels of EARLI1 and COR15α steady-state mRNA. Upper two panels: EARLI1 is transiently induced by cold, but not by ABA. ABA treatment: late-flowering FRIGIDA-containing Columbia seedlings (Col[FRI]) grown in vitro for 7 days at room temperature in short-day photoperiods were flooded with 10 ml of 100 μM ABA for 5 min, then washed with sterile water. RNA was prepared from seedlings grown for the indicated length of time in hours (Hrs) after the ABA treatment. Transient cold treatment: late-flowering Col(FRI) seedlings were grown in vitro for 7 days at room temperature in short-day photoperiods, then transferred overnight to 4°C. RNA was prepared from seedlings grown for the indicated length of time in hours (Hrs) at room temperature after the cold treatment. A full-length cDNA probe was used to detect EARLI1 mRNA and a 3' UTR-specific cDNA fragment of the actin gene ACT2/ACT8 (An et al., 1996) to determine the relative amount of RNA in each lane. Lower two panels: the cold-regulated gene COR15α is transiently induced by both cold and ABA. Col(FRI) seedlings were treated as described above. A3'-UTR-specific cDNA fragment were used to detect COR15α mRNA (Baker et al., 1994). The actin gene ACT2/ACT8 was used to determine the relative amount of RNA in each lane. It may be worth noticing that ABA lowers the level of actin steady-state mRNA.